

CD34⁺VEGFR-3⁺ progenitor cells have a potential to differentiate towards lymphatic endothelial cells

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Introduction

Lymphatic vessels

no basement membrane

no pericytes

functions:

- draining excess fluid from extracellular space
- absorbing & transportation of lipids
- transporting leukocytes & APC
- removal of cell debris, dust particles & microorganism
- tumour metastasis

Lymphangiogenesis

occurs in

- wound healing
- embryonic development
- tissue regeneration
- inflammation
- tumour metastasis

*newly formed lymphatic vessels
sprout from pre-existing vessels by
proliferation, migration and tube
formation of endothelial cells*

Introduction

Endothelial progenitor cells

are circulating cells that promote neovascularization at sites of ischemia, injury, hypoxia or tumour formation

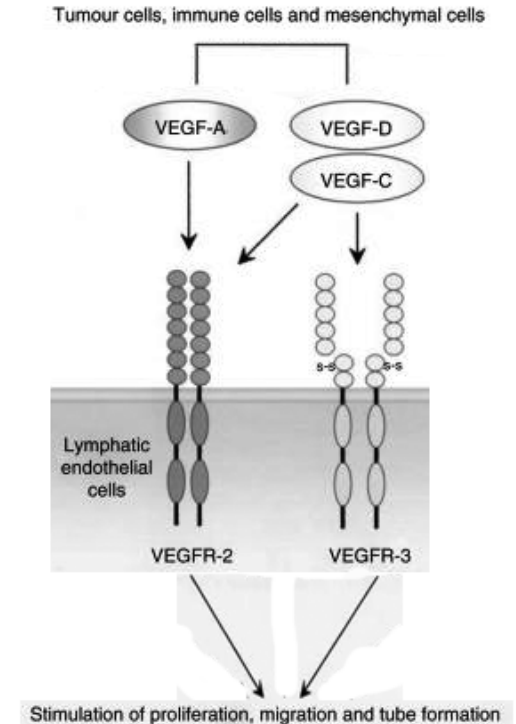
majority resides in bone marrow

number of EPCs in human cord blood is much higher than in peripheral blood

were already detected at growing lymphatic vessels in the cornea of mouse & transplanted human kidney

Introduction

- CD34 haematopoetic cells, EPCs
- CD133 haematopoetic stem/progenitor cell marker
- VEGF-C
- VEGFR-3 receptor of VEGF-C/-D , expressing specifically on lymphatic endothelial
- Prox-1 a lymphatic specific marker expressed in nuclei
- 5'Nase marker for lymphatics
- LYVE-1 marker or lymphatics, may play a role in lymphatic hyaluronan transport
- CD44 is a receptor for hyaluronic acid

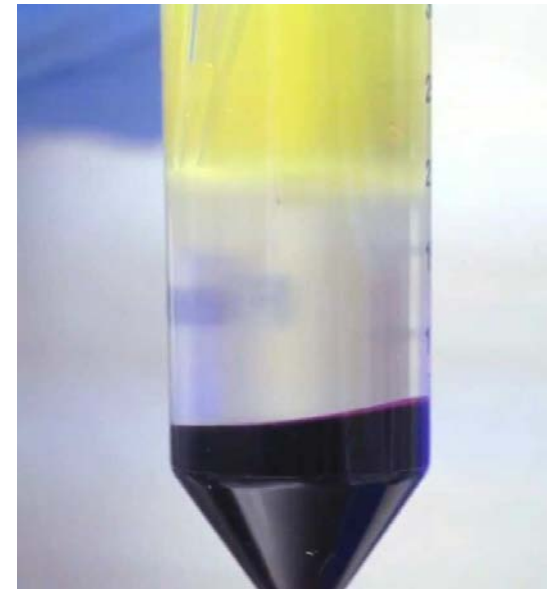


Harvesting of mononuclear cells

40-60 ml human umbilical cord blood

mononuclear cell isolation by density-gradient centrifugation (percoll)

examination of cells under confocal laser microscope



Isolation & identification of EPCs

detection of CD34⁺VEGFR-3⁺ and CD133⁺VEGFR-3⁺ cells by fluorescence-activated cell sorting

examination of CD133 expression on CD34⁺VEGFR-3⁺ cells

evaluation of uptake of acetylated low-density lipoprotein (Dil-Ac-LDL) and Ulex Europaeus agglutinin-1 (UEA-1)

Induction of cell differentiation

seeding of CD34⁺VEGFR-3⁺ cells on dishes (pre-coated with fibronectin)

incubation in medium enriched with VEGF-C for at least 14 days

observation of morphological changes at day 1, 7, 10, 14 & 21

for following experiments only cells which were incubated in VEGF-C enriched medium for 14 days were used

Immunocytochemistry

to evaluate if VEGF-C induced EPCs differentiated towards lymphatic endothelial cells, they were seeded on coverslips (pre-coated with polylysine) and stained for

5'Nase

LYVE-1

Prox-1

CD44

Quantitative real-time PCR

extraction of total RNA from CD34⁺VEGFR-3⁺ cells treated with VEGF-C at different time-points

determination of VEGFR-3 mRNA level

control: CD34⁺VEGFR-3⁺ cells without VEGF-C treatment

RNA interference

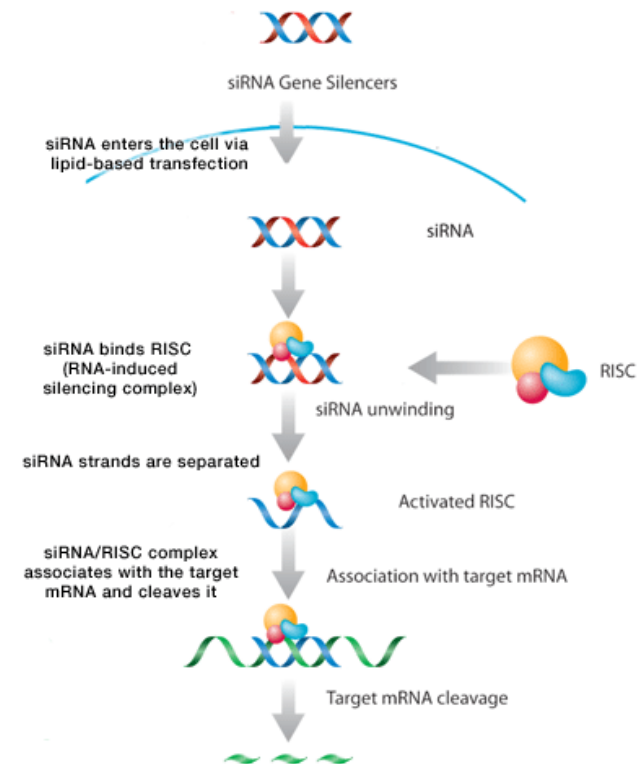
to evaluate the effect of VEGF-C/VEGFR-3 pathway
on lymphangiogenesis

3 different siRNAs targeting human VEGFR-3 and 1
irrelevant siRNA (control) were designed

transfection of cells with Lipofectamine 2000 →

incubation in VEGF-C enriched medium

assessment of VEGFR-3 mRNA levels by RT-PCR



4 groups

vehicle

VEGF-C

VEGF-C + VEGFR-3 siRNA

VEGF-C + irrelevant siRNA

Proliferation assay

cells were seeded on fibronectin pre-coated dishes

after 24 h incubation cells were counted with a haemocytometer

assays were repeated 5 times

4 groups

vehicle

VEGF-C

VEGF-C + VEGFR-3 siRNA

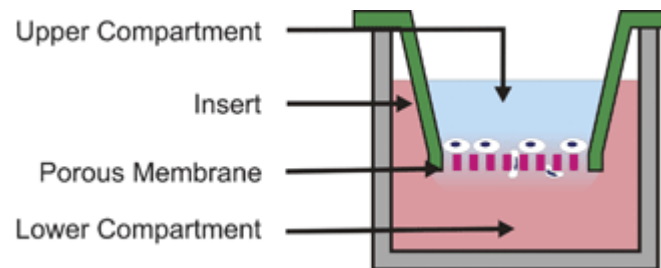
VEGF-C + irrelevant siRNA

Transmigrating assay

VEGF-C was placed in lower chamber

after 12 h incubation, cells in the lower chamber were counted

assays were repeated 3 times



Wounding assay

cells in dishes were scraped on 4 different sides

cells were counted in successive 100 μm section of 300 μm wide

maximal distance of cell migration from the wounded edge was measured

assays were repeated 3 times

5 groups

vehicle

bFGF

VEGF

VEGF-C

VEGF-C + VEGFR-3 siRNA

4 groups

vehicle

VEGF-C

VEGF-C + VEGFR-3 siRNA

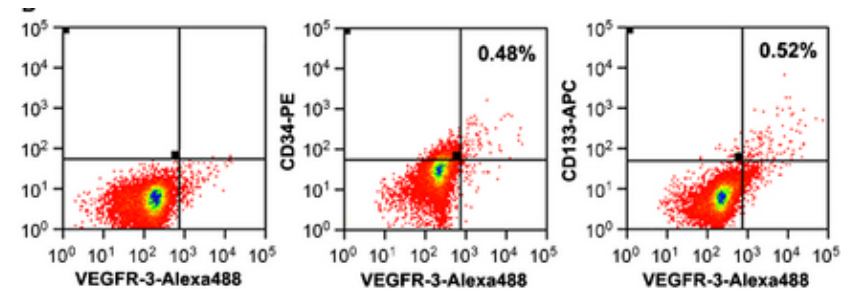
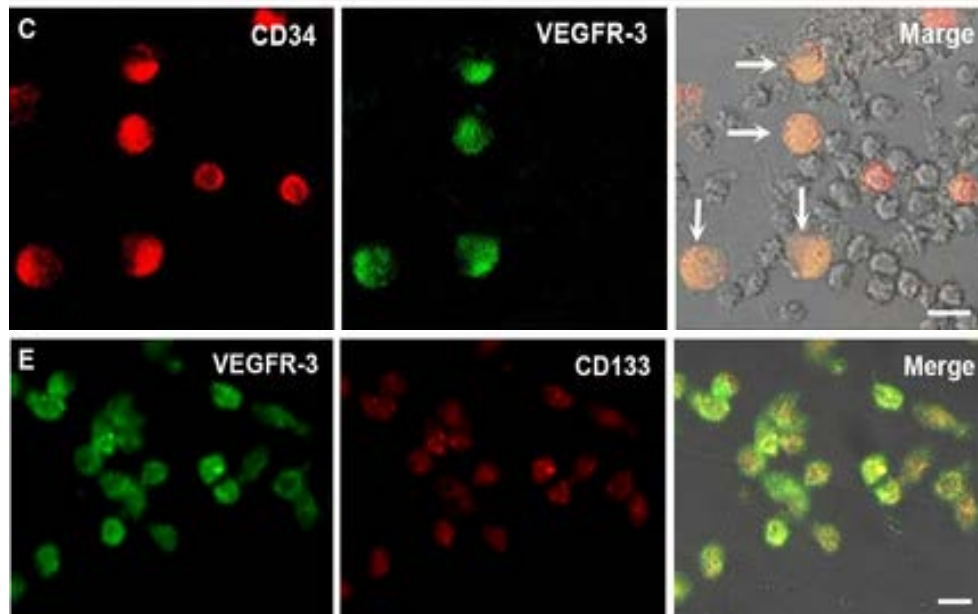
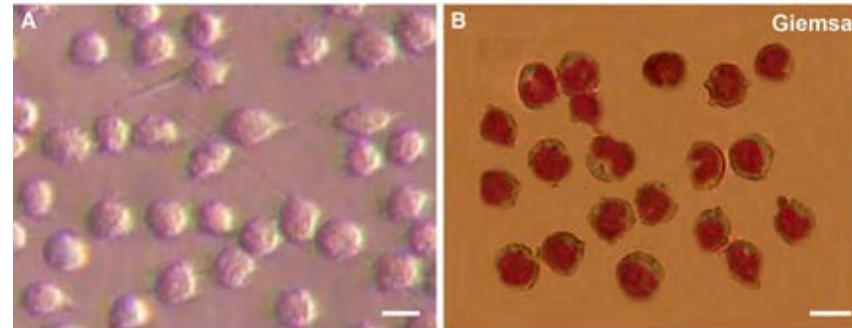
VEGF-C + irrelevant siRNA

Tube formation in 3D collagen gel & Matrigel

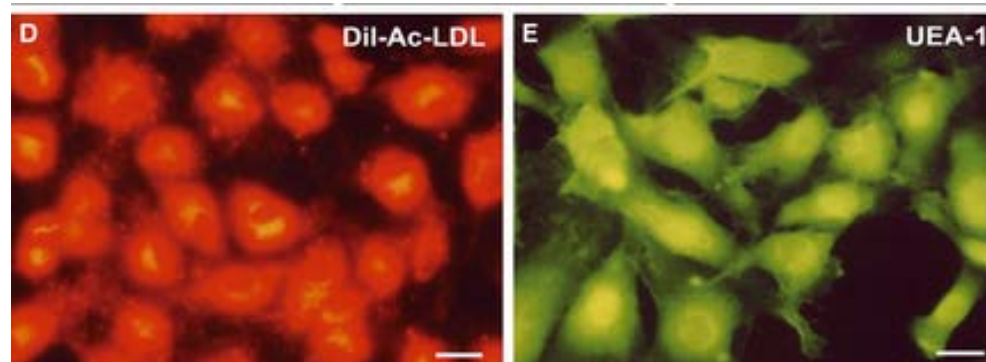
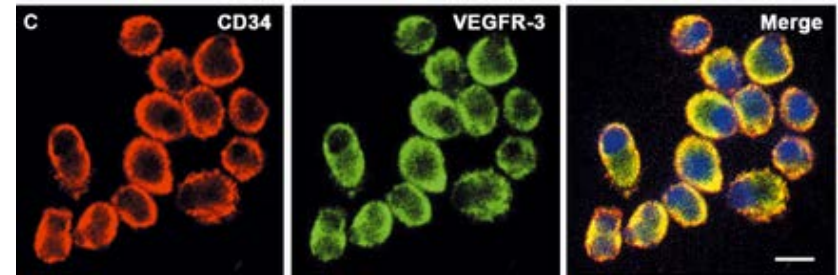
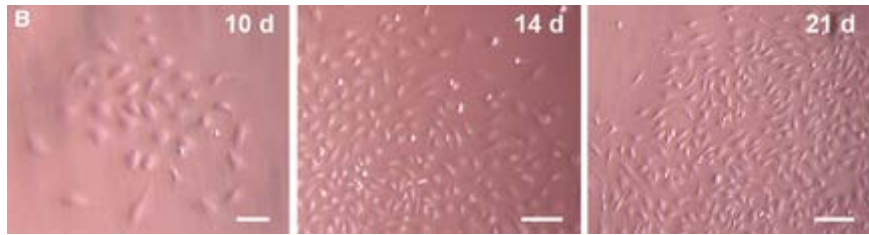
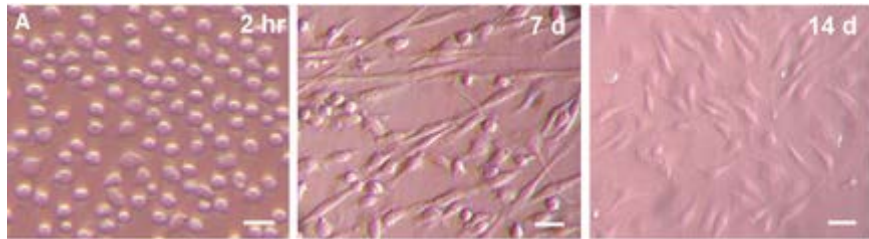
collagen gel: collagen type I from rat tail, dissolved in acetic acid
DMEM
NaHCO₃

cells were seeded on dishes (pre-coated with collagen), when cells grew to monolayer they were treated and afterwards a layer of collagen was made
tube formation & area + length of tubes were assessed using TEM
LYVE-1 staining for identifying as lymphatic capillary like structures
assays were repeated 3 times

Results

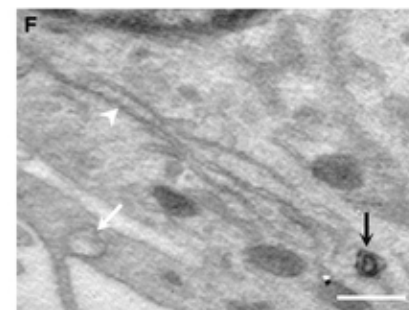
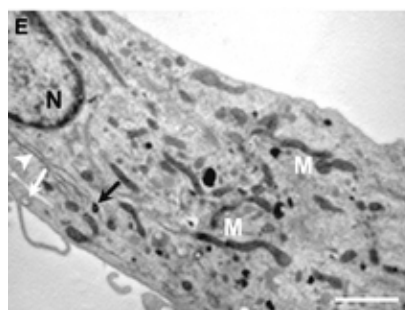
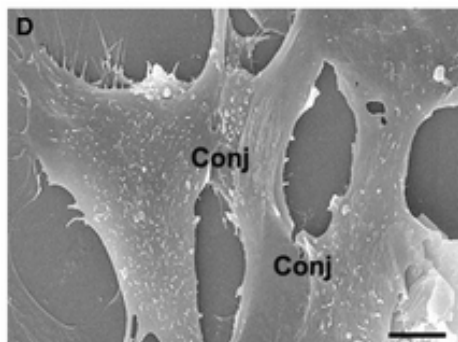
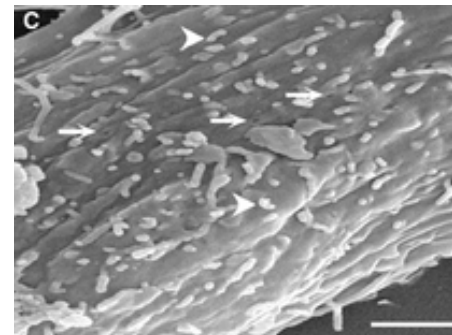
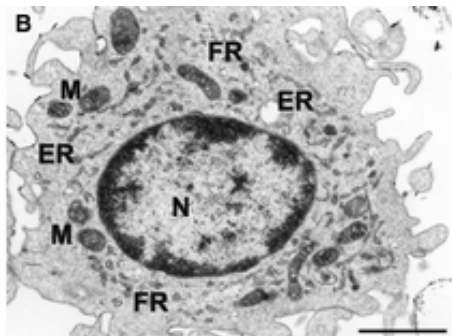
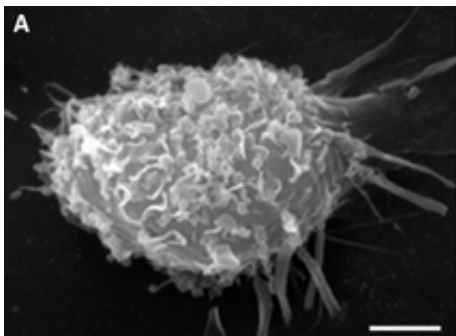


Results

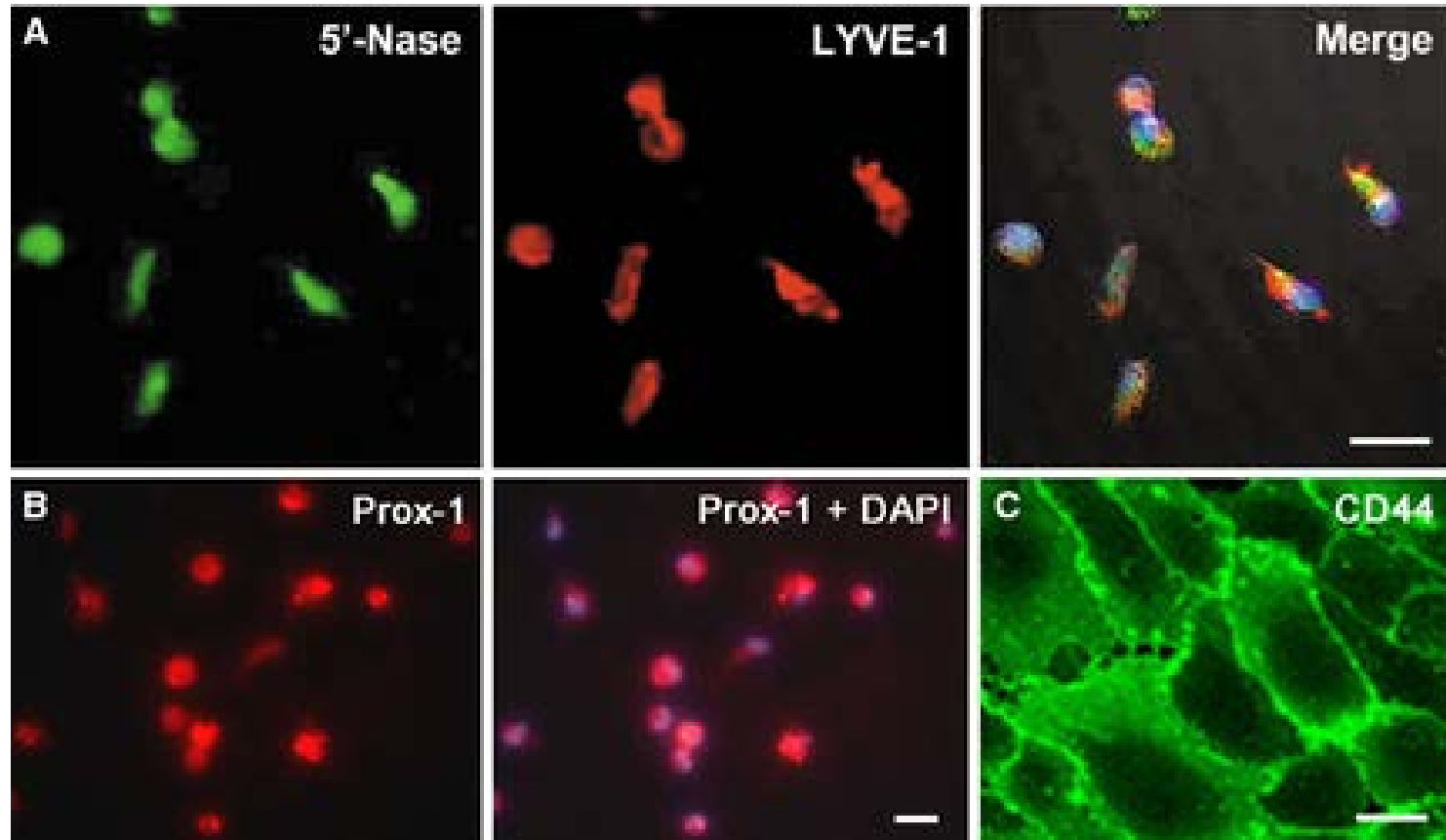




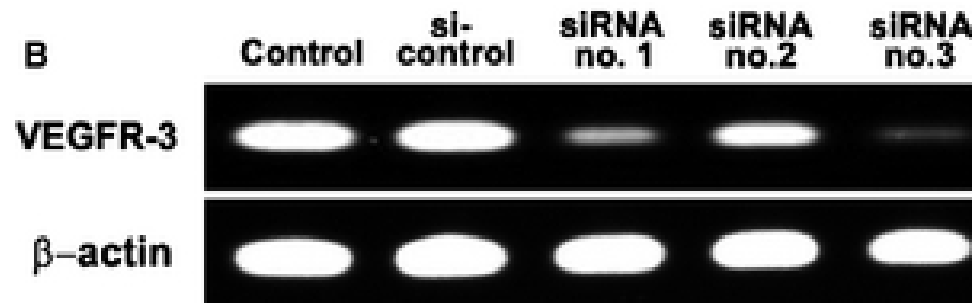
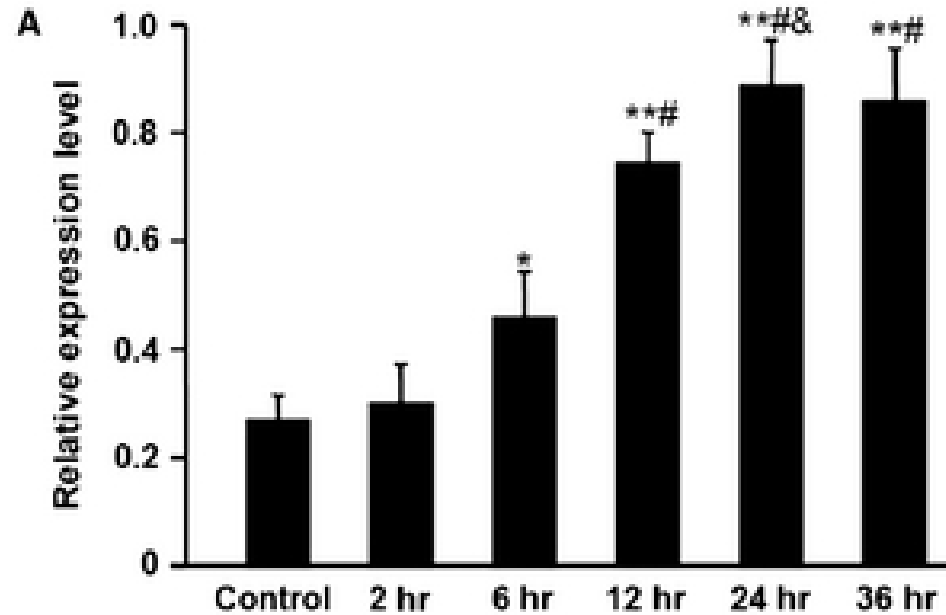
Results



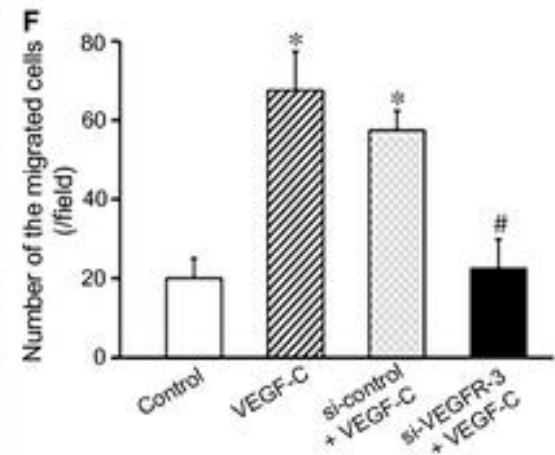
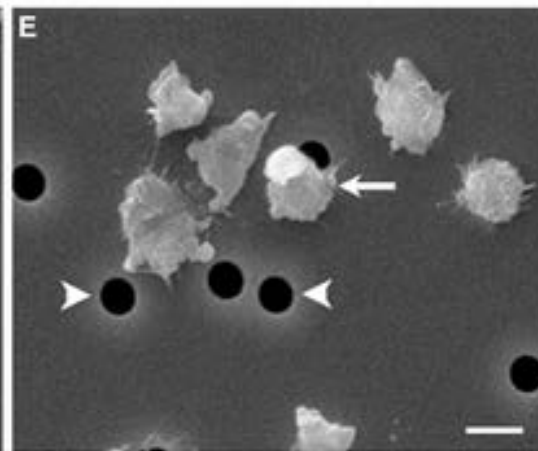
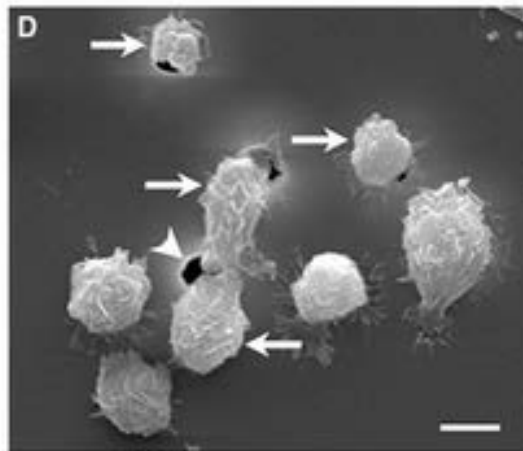
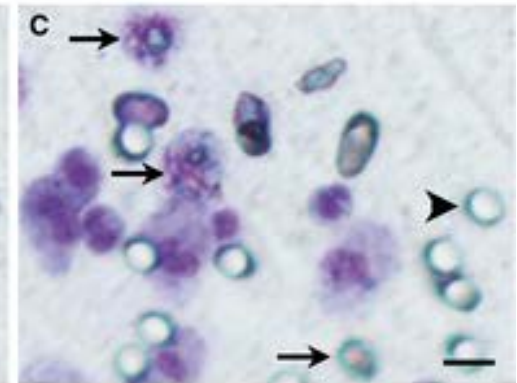
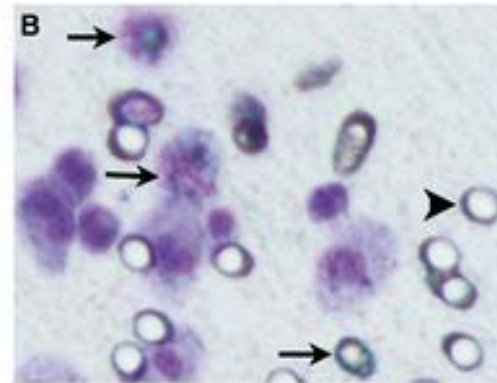
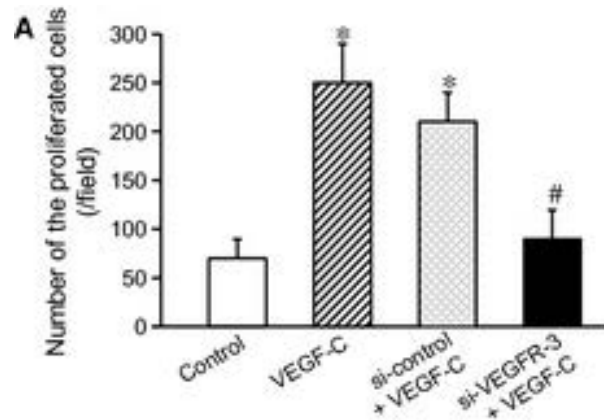
Results



Results



Results



Results

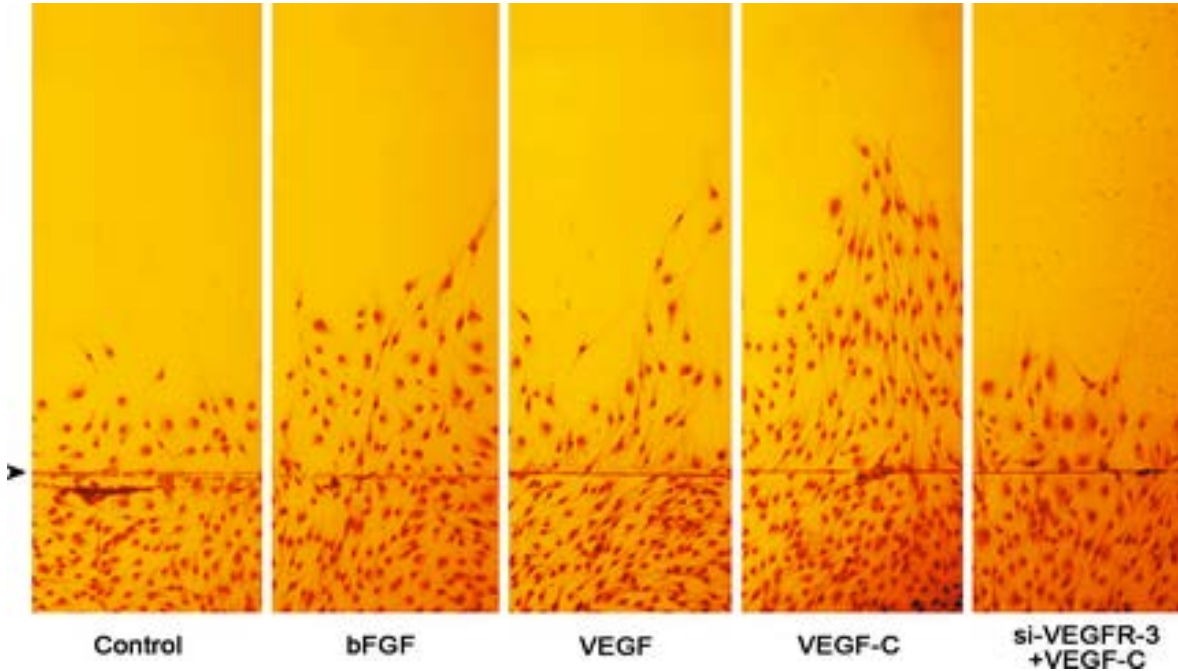


Table 1 Effects of bFGF, VEGF and VEGF-C on migration of the EPC-derived cells

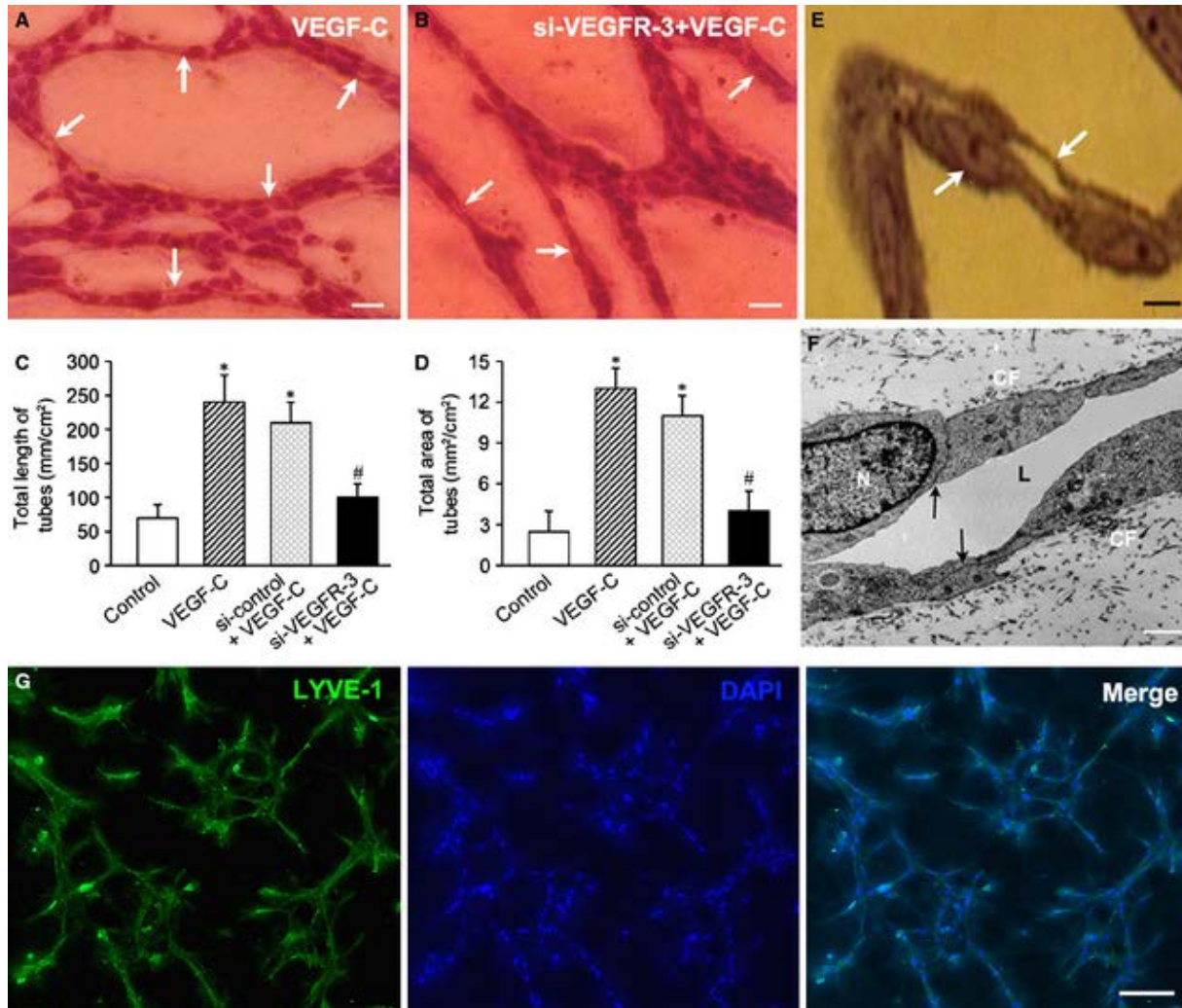
Groups	Cell numbers	Maximal distance (μm)
Control	14 \pm 3	239 \pm 36
bFGF	36 \pm 8*	398 \pm 28*
VEGF	30 \pm 6*	386 \pm 42*
VEGF-C	48 \pm 10* [†]	578 \pm 48* [†]
RNAi	16 \pm 5 [#]	242 \pm 39 [#]

The values are mean \pm SD.

* $P < 0.01$ versus control group, [†] $P < 0.01$ versus bFGF and VEGF groups, [#] $P < 0.01$ versus VEGF-C group. $n = 16$.



Results



Conclusion

VEGFR-3⁺CD34⁺ cells were found & isolated in human cord blood

VEGF-C induction lead to differentiation into LECs

proliferation of LECs

migration of LECs

LECs could form tubes which connected and formed a network in 3D collagen gel

VEGF-C/VEGFR-3 signalling pathway may be crucial in lymphangiogenesis