



# Productive HIV-1 infection of human cervical tissue *ex vivo* is associated with the secretory phase of the menstrual cycle

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HIV-1 deposited in mucosa of lower female reproductive tract (FRT)

Infects primary HIV-1 target cells and disseminates to the regional lymph nodes

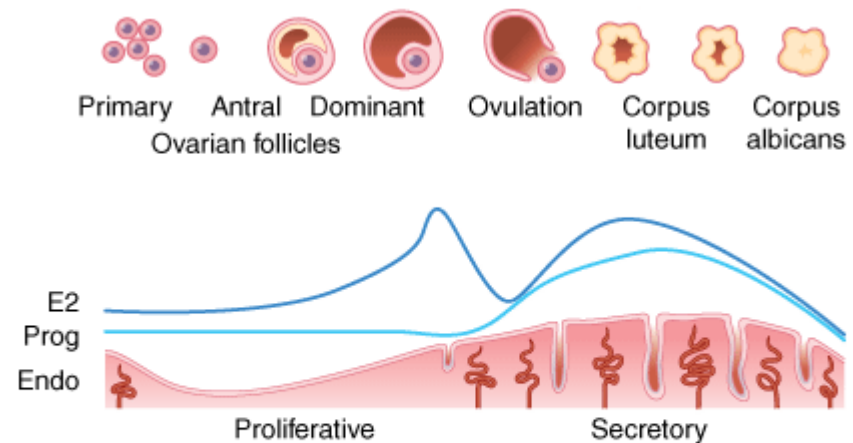
Challenge: Lack of understanding of basic mechanisms of HIV-1 transmission + dissemination in FRT

Protection against sexually transmitted infections (STIs) versus enabling successful reproduction.

Estradiol + progesterone modify functions of epithelial cells, fibroblasts and immune cells in the FRT

Innate and adaptive immune systems under hormonal control  
-> immune protection in FRT varies with phase of menstrual cycle.

Immune protection dampened during secretory phase of cycle to optimize conditions for fertilization and pregnancy  
-> "Window of Opportunity" for pathogens



Source: Fauci AS, Kasper DL, Braunwald E, Hauser SL, Longo DL, Jameson JL, Loscalzo J: *Harrison's Principles of Internal Medicine*, 17th Edition: <http://www.accessmedicine.com>  
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# Mucosal Barrier

**Immune function**

**Effects of oestradiol and progesterone**

**Lumen**

Pathogen killing and inactivation

**Lower FRT**

↓ HBD2, elafin, IgA and IgG

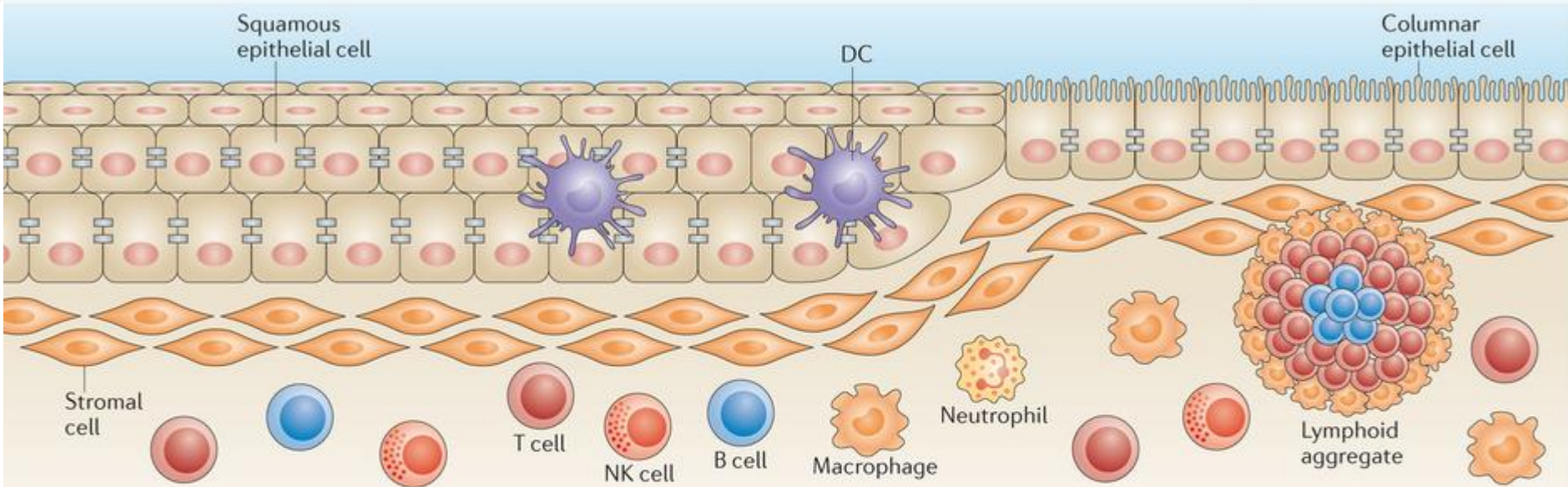
**Upper FRT**

↑ CCL20, HBD2, SLPI, elafin, IgA and IgG (pIgR and FcRn)

Squamous epithelial cell

DC

Columnar epithelial cell



Stromal cell

T cell

NK cell

B cell

Macrophage

Neutrophil

Lymphoid aggregate

**Epithelial cells**

Immune cell recruitment, innate immune protection and tolerogenic nature

↓ CCL2, IL-8, HBD2 and elafin

↑ CX<sub>3</sub>CL1, CCL2, IL-8, HBD2, CCL20, SLPI, elafin and TGFβ

**Fibroblasts**

Immune cell recruitment

↔ CXCL12

↑ CXCL12

Regulation of epithelial cells

↔ HGF

↑ HGF

**Immune cells**

Pathogen killing

↔ CD8<sup>+</sup> CTL activity

↓ CD8<sup>+</sup> CTL activity

Protection against viruses

↔ NK cell activity (IFNγ)

↓ NK cell activity (IFNγ)

Protection against bacteria

CD14<sup>+</sup> macrophages

CD163<sup>+</sup> macrophages

Infection by HIV

↓ CD4<sup>+</sup> T cell susceptibility

↓ CD4<sup>+</sup> T cell susceptibility

## Cervico-vaginal tissue explants (CTE)

Retain the *in vivo* cyto-architecture and some tissue functions for several days

Previous CTE experiment: activated tissue-associated CD4+ T cells are major targets ->

likely source of CCR5-dependent (R5), but not CXCR4-dependent (X4), HIV-1

Contribution of resident macrophages needs more research

CTEs obtained from HIV-seronegative women undergoing hysterectomy for benign gynecological conditions.

# R5 HIV-1 replication

CTEs dissected into  $\sim 2\text{-mm}^3$  blocks, cultured on collagen sponge gel rafts for up to 12 days

CTEs inoculated with R5 HIV-1<sub>BaL</sub>

Virus replication evaluated by p24<sub>Gag</sub> release into culture supernatant and by accumulation of HIV DNA in tissue-associated cells.

Control donor-matched tissue blocks incubated with R5 HIV-1 in presence of the RT inhibitor 3TC

# R5 HIV-1 replication

Virus replication evident on day 9 post-HIV-1 inoculation

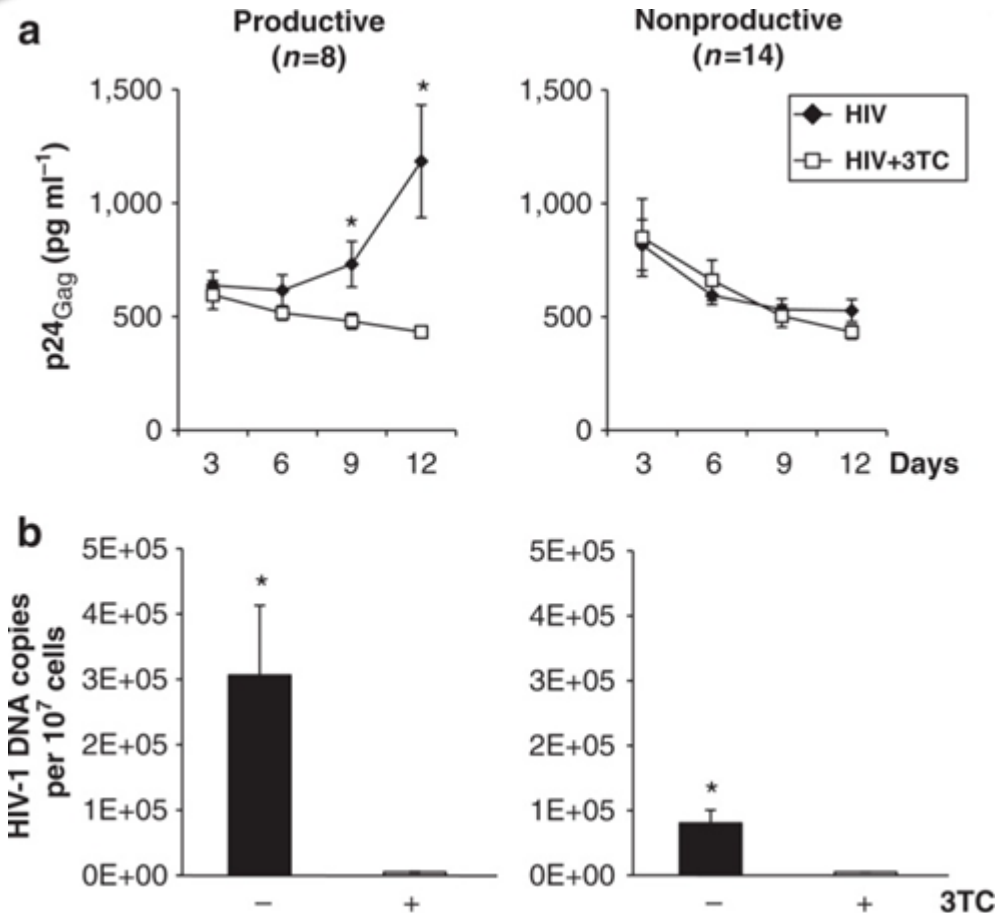
Increased up to day 12 as evaluated by p24<sub>Gag</sub> release in CTE culture supernatants

No increase of virus release in 3TC-treated HIV-1-exposed cultures

“**Productive**”: CTEs with progressive accumulation of p24<sub>Gag</sub> into culture supernatants (n=8)

“**Nonproductive**”: CTEs showing no difference in p24<sub>Gag</sub> release in the presence or absence of 3TC (n=14)

# R5 HIV-1 replication





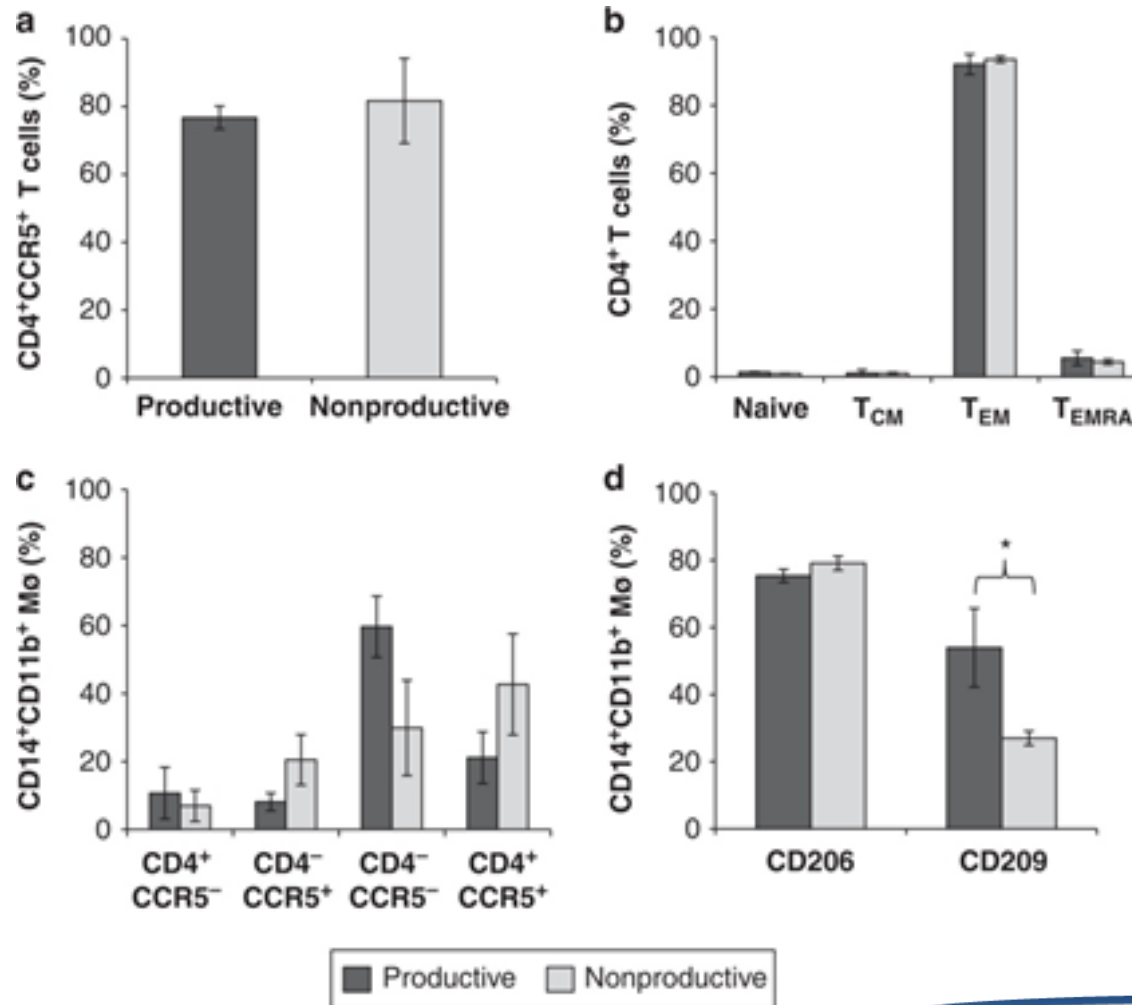
CTEs digested with collagenase IV

Cell suspensions stained for different surface antigens

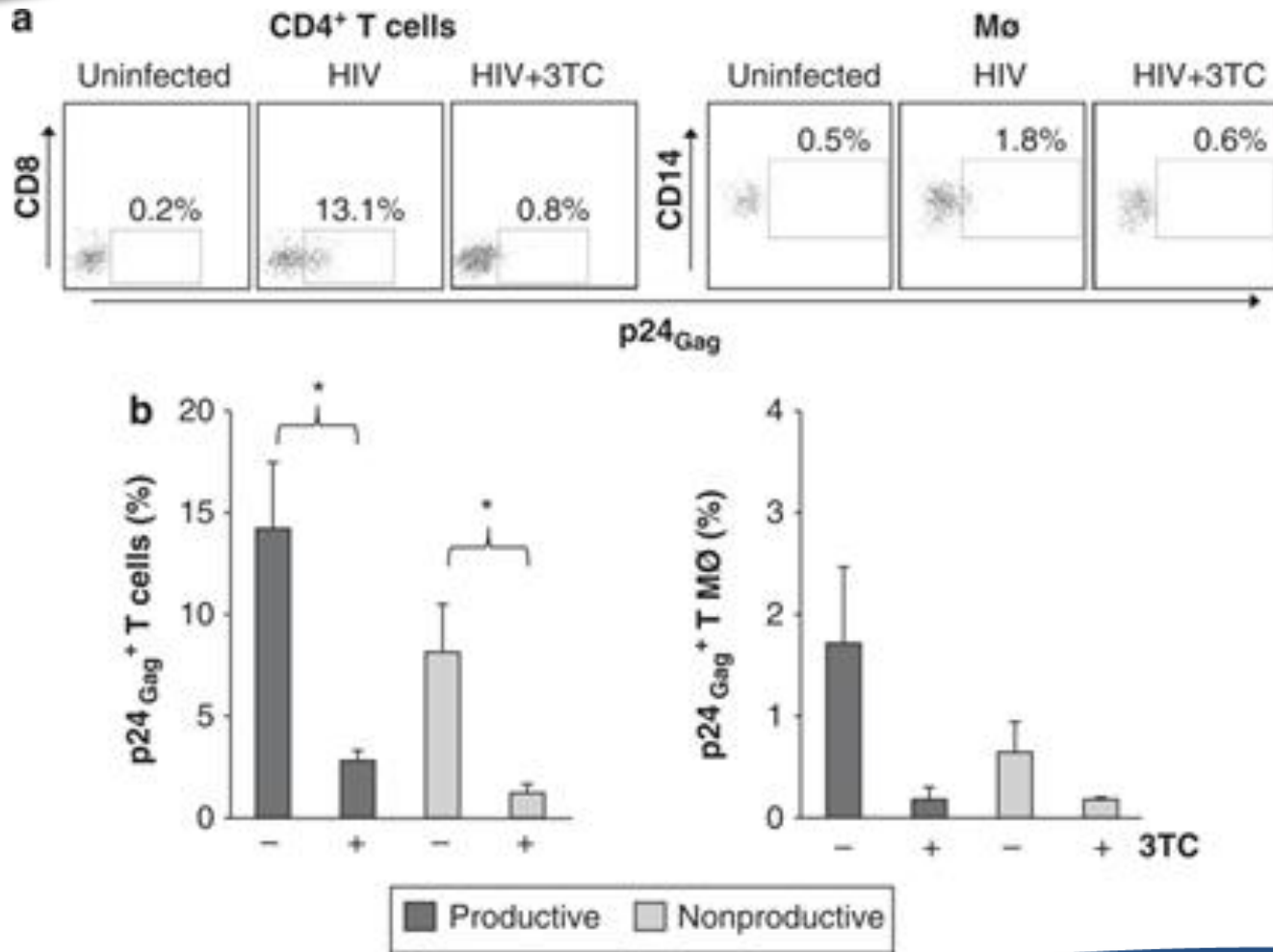
Flow cytometric analysis -> % of CD4+ T cells and resident macrophages not significantly different between productive and nonproductive tissues

Most CD3+CD4+ T cells were CCR5+ and showed an “effector memory” phenotype, as defined by lack of expression of CCR7 and CD45RA

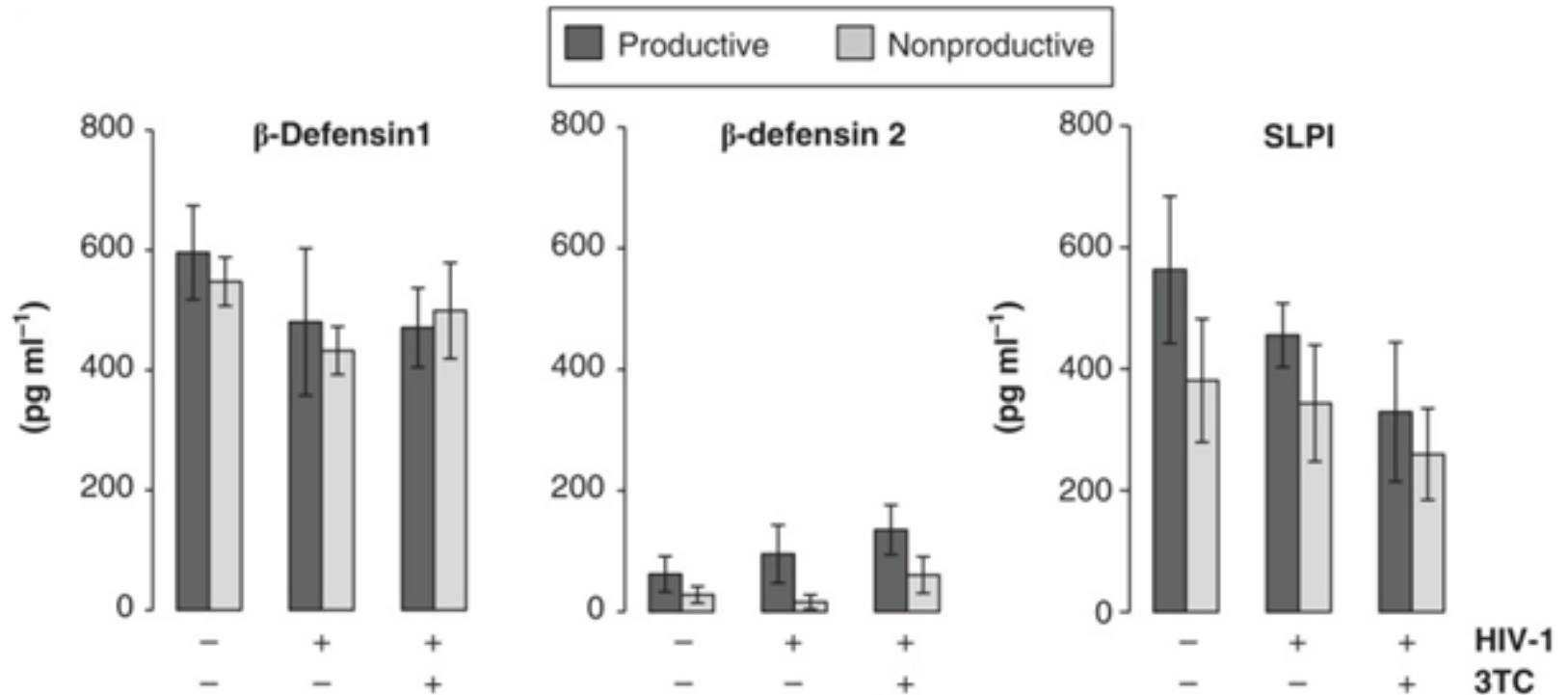
# Susceptibility to HIV-1 infection of CD4+ T cells and macrophages



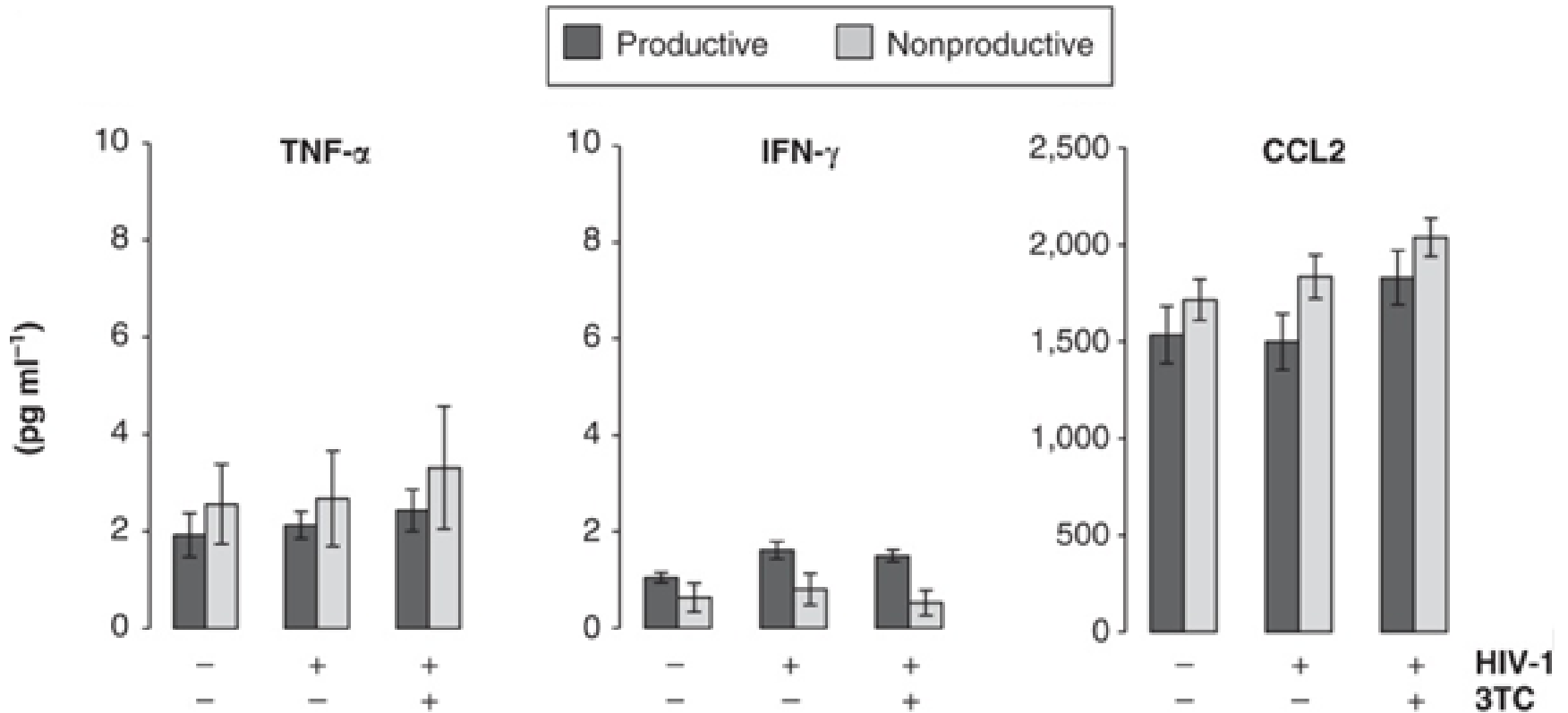
# Infection of HIV-1 target cells



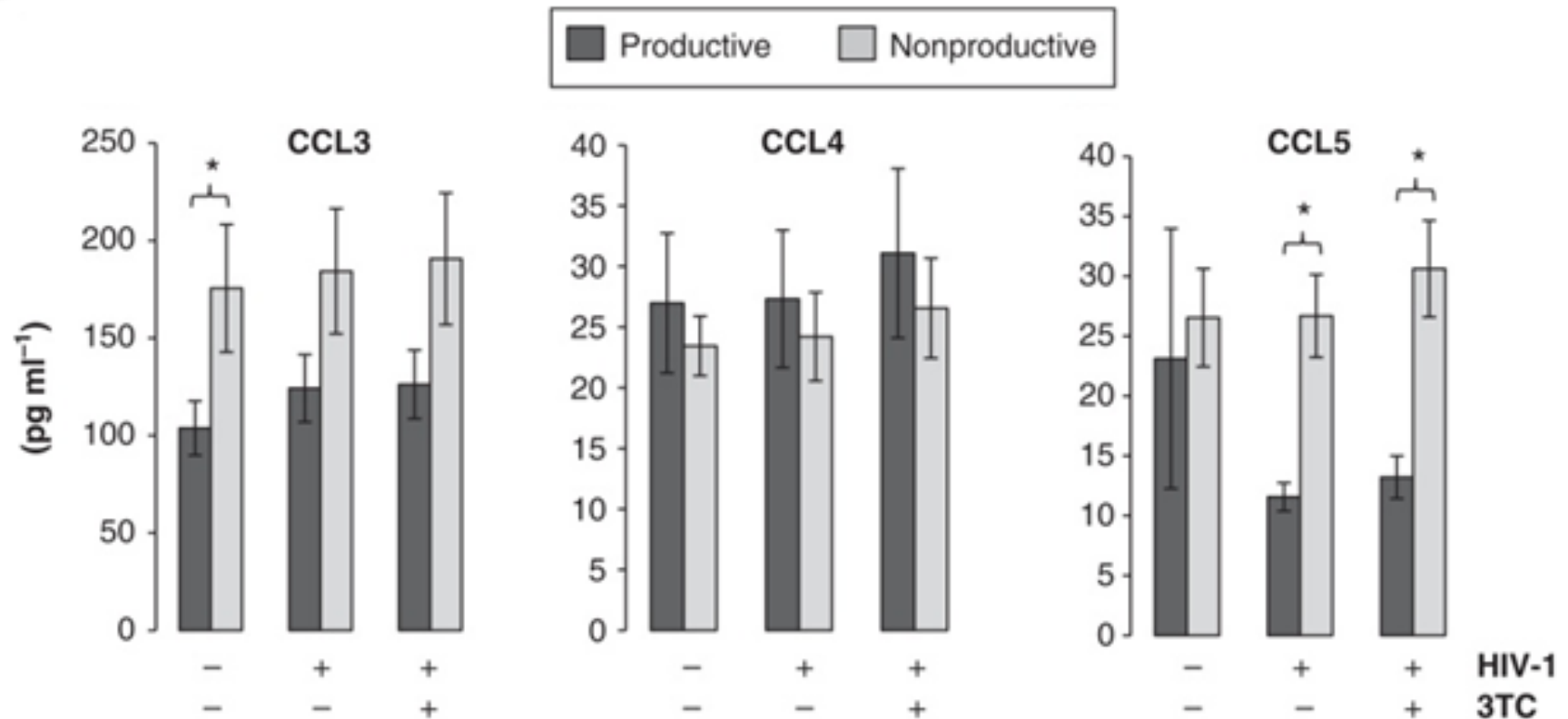
# Anti-viral peptides



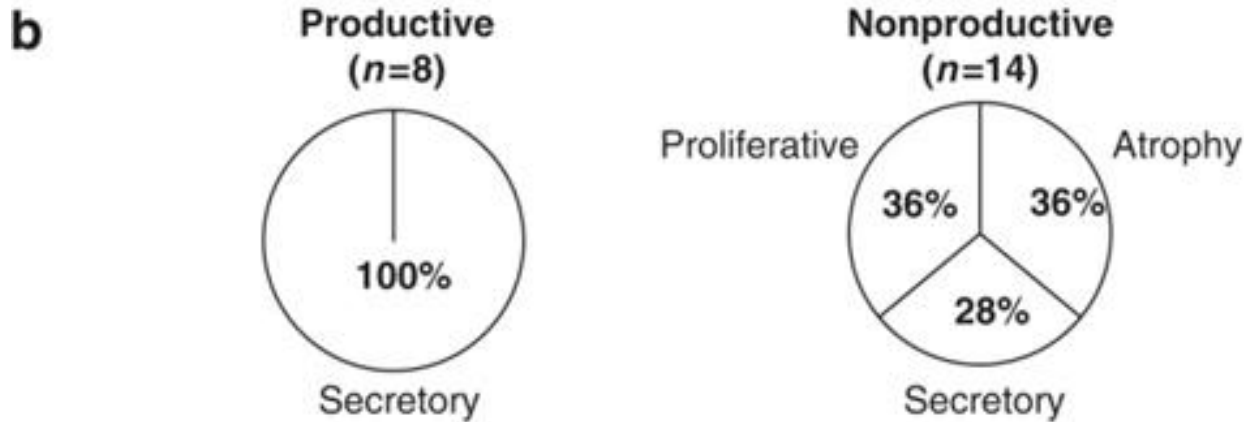
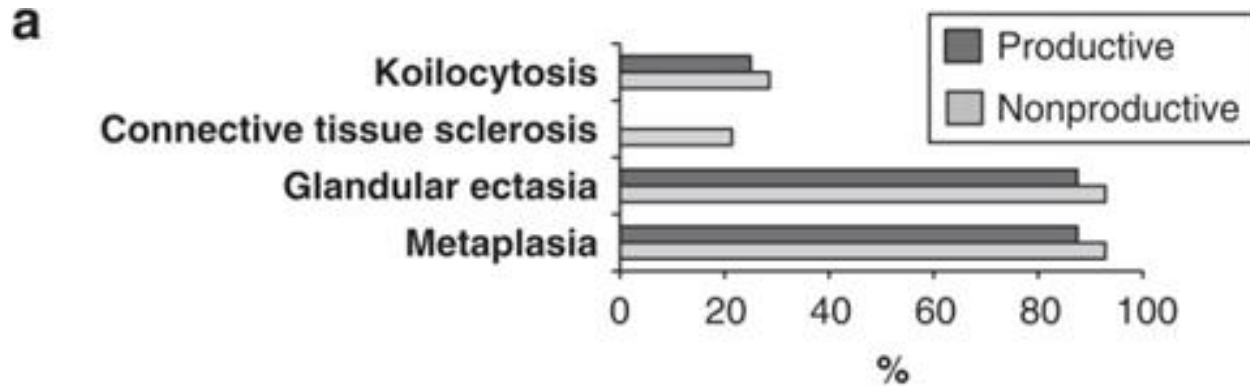
# Pro-inflammatory cytokines



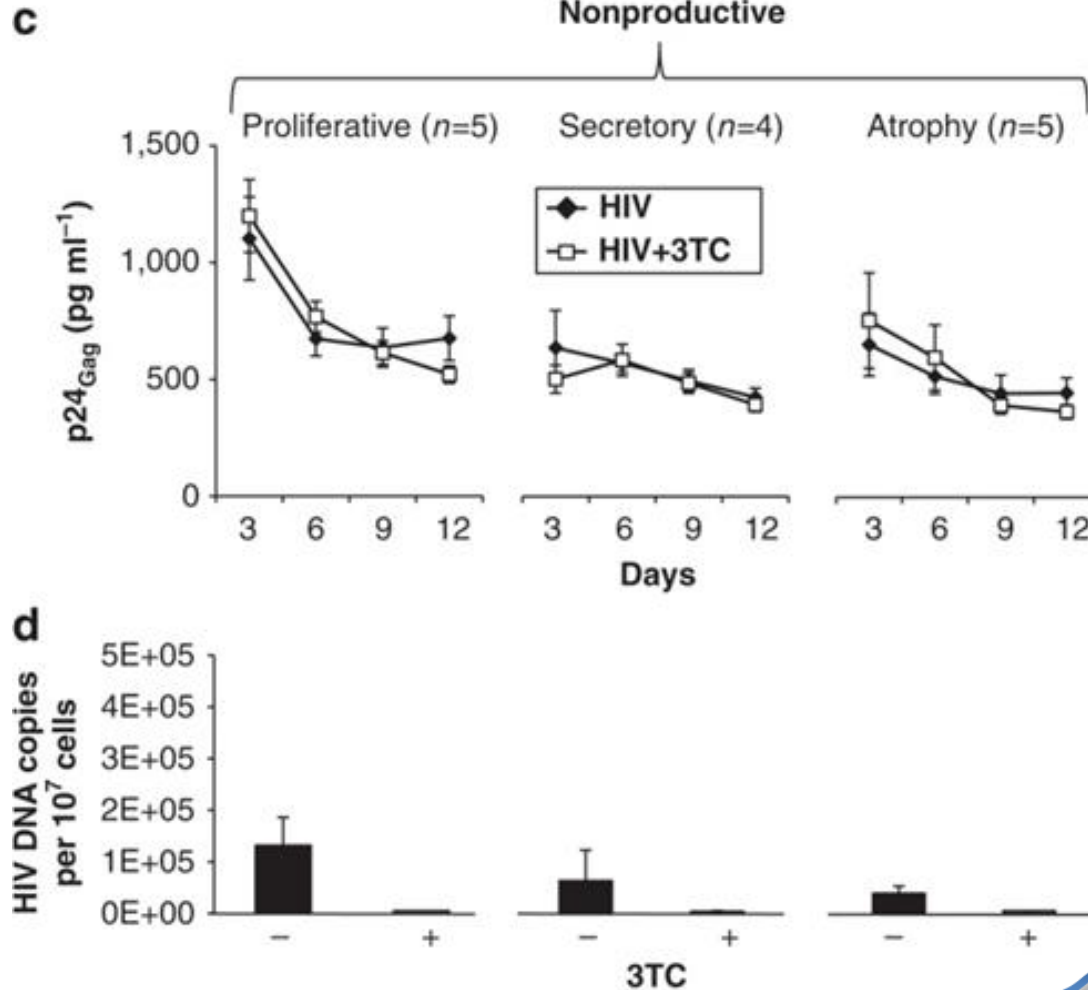
# CCR5 ligands



# Productive and nonproductive CTE tissue architecture



# Productive and nonproductive CTE tissue architecture





Investigated susceptibility to productive R5 HIV-1 infection of CTEs *ex vivo* from 22 HIV-1 seronegative women

8 out of 22 CTEs (36%) showed virus replication

14 out of 22 CTEs (64%) in presence or absence of 3TC did not

-> “productive” and “nonproductive” infections.

Both CD4+ T cells and resident macrophages supported HIV-1 replication, although macrophages to lesser extent than T lymphocytes

Outcome infection (productive vs. nonproductive) did not correlate with number of HIV-1 target cells

# Summary

Functional distributions of CD4<sup>+</sup> T cells similar in productive and nonproductive CTEs, with CD4<sup>+</sup> T cells displaying an “effector memory” phenotype representing the dominant subset

Resident macrophages: % expressing CD206 (mannose receptor) comparable in both productive and nonproductive CTEs but significantly higher % of cells expressing CD209 (DC-SIGN) on surface in productive CTEs than in nonproductive ones

No significant differences in secreted levels of innate anti-viral peptides ( $\beta$ -defensins, SLPI), pro-inflammatory (TNF- $\alpha$ , IFN- $\gamma$ , CCL2), or anti-inflammatory (IL-4, IL-10) cytokines between productive and non-productive

Some CCR5-binding chemokines secreted at higher levels by nonproductive vs. productive CTEs

# Summary

*Post-hoc* analysis of *ex vivo* outcomes of HIV-1 exposure of CTEs (productive vs. nonproductive) did not reveal a correlation with histological alterations of mucosa and submucosa.

Strong correlation between productive HIV-1 infection of CTEs and menstrual cycle phase of the donor at the time of surgery

-> infected CTEs all from women in secretory phase, nonproductive tissues from women in different phases of menstrual cycle

Menstrual cycle of women and/or their consumption of hormonal-based contraceptives should be taken into consideration when new preventative strategies against HIV transmission are developed

CTEs can represent an adequate experimental system to study the mechanisms and replication of HIV-1 infection

CTEs could be adopted as low-cost platform for testing novel antivirals and microbicides before moving towards more expensive NHP models of HIV-1 transmission and clinical trials in women.

Thank you for your attention!