

Mesenchymal stem cells release exosomes that transfer miRNAs to endothelial cells and promote angiogenesis

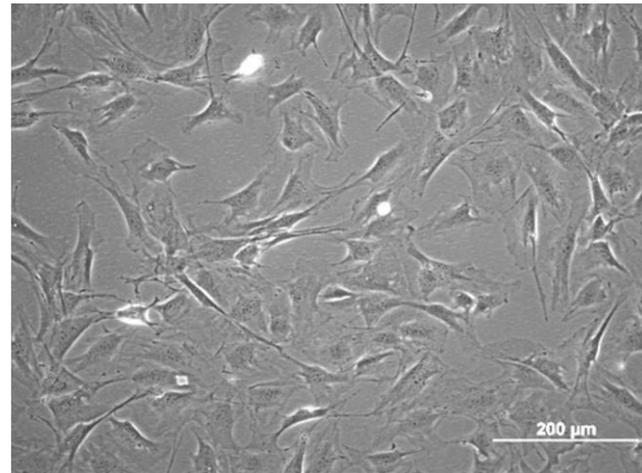
Gong *et al.*, *Oncotarget*. 2017 Apr 1.

Tanja Wagner

Introduction

Mesenchymal stem cells (MSCs)

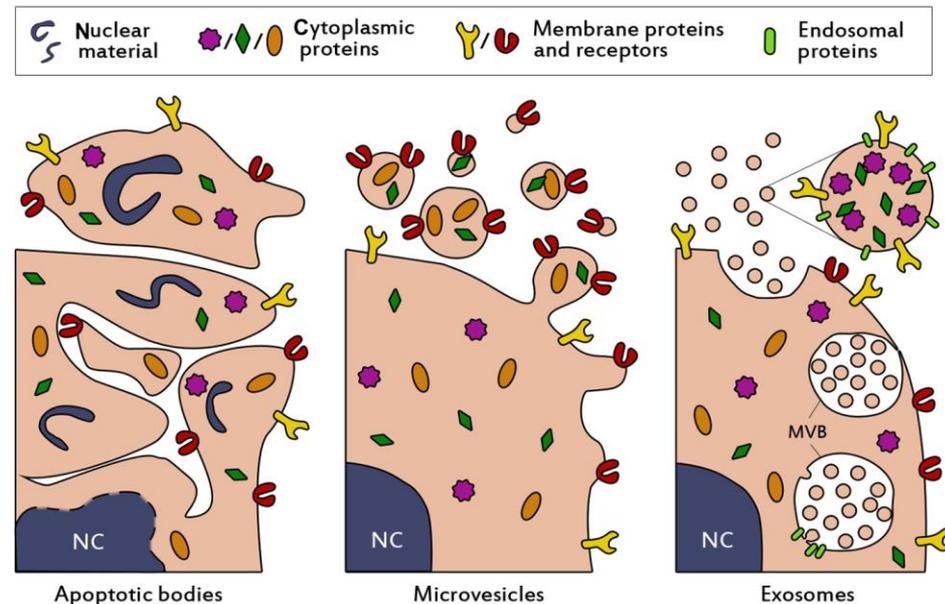
- non-haematopoietic, multipotent stem cells with the capacity to differentiate into mesodermal lineage such as osteocytes, adipocytes and chondrocytes as well ectodermal (neurocytes) and endodermal lineages (hepatocytes)
- Have a spindle-shaped fibroblast like morphology
- Can increase endothelial cell growth and enhance new blood vessel formation



Gong *et al.*, 2017

Exosomes

- Cell-derived vesicles: diameter 30-100 nm
 - Originate from budding into the limiting membrane of large endosomal structures (multivesicular bodies =MVB) in the cytosol
- MVB are able to fuse with the plasma membrane, causing the release of exosomes into the extracellular space



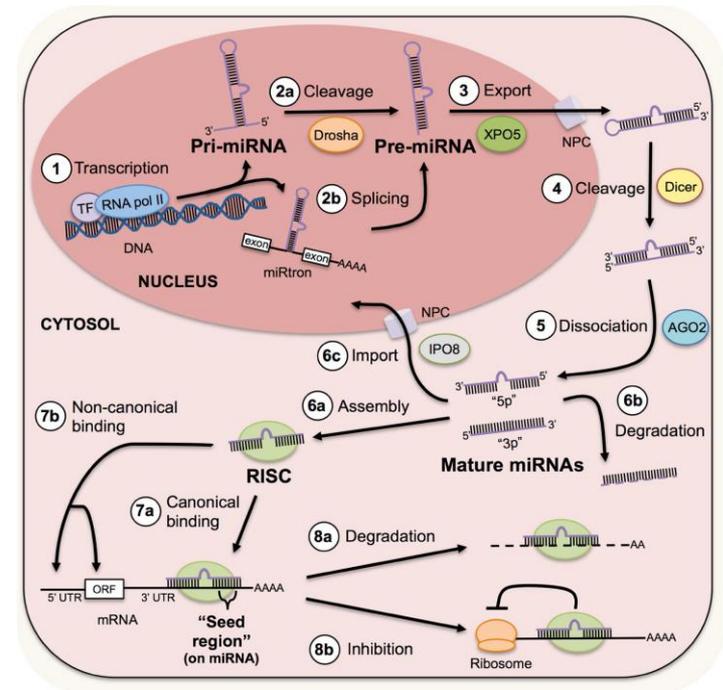
Kooijmans *et al.*, 2012

Exosomes

- Exist in almost **all biological fluids** including blood, urine, saliva, cerebrospinal fluid, and cell preconditioned medium
- Shuttle **mRNAs, miRNAs** and **other molecular constituents** to achieve **cell-to-cell communication**

miRNAs

- Small non-coding RNAs (containing about 18-22 nucleotides)
- Regulate gene expression on the **post-transcriptional level** by binding to specific mRNA and inducing their
 - degradation
 - translational inhibition
- Play a role in biological and pathological processes including the cell cycle, hematopoiesis, neurogenesis, aging, cancer and cardiovascular diseases
- miR-30 family targeted DLL4 in endothelial cells to promote angiogenesis



Gerlach and Vaidya, 2017

Hypothesis

Whether MSC-derived exosomes shuttle various pro-angiogenic miRNAs and transfer these miRNAs to endothelial cells resulting in promoting angiogenesis

Methods

Conditioned medium derived from MSCs

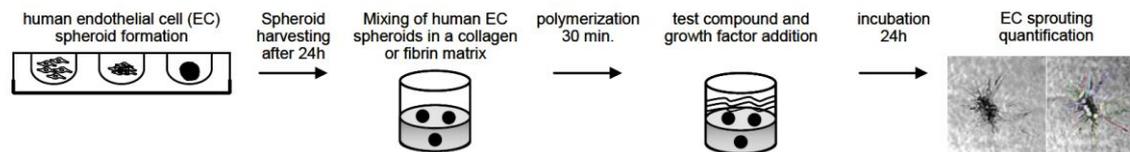
1. MSCs cultured in complete DMEM/F12 medium for 24h
2. Medium was replaced with 15 ml of serum-free medium
3. After 48 h culture the medium was collected and centrifuged to remove cell debris
4. Supernatant was filtered and centrifuged at 3200g at 4°C for 45 minutes
5. Transferred into ultra-filtration conical tubes to concentrate medium to 100x
6. **Exosomes** were isolated from concentrated CdM using an ExoQuick-TC Exosome Precipitation Solution
7. **Exosome pellets** were resuspended with DMEM medium and stored at -80°C

Angiogenesis models

1. Tube-like structure formation assay

- HUVECs were seeded on top of Matrigel
- Treated with CdM or exosomes (100 µg/ml) for 16h
- Images were taken

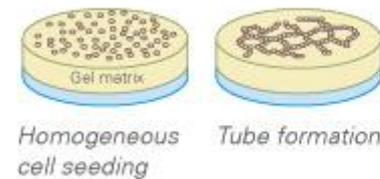
2. Spheroid-based sprout assay



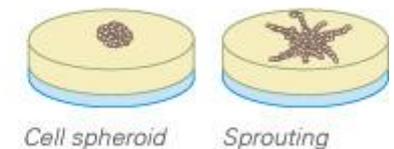
www.proqinase.com

- GFP+ HUVECs (500cells/spheroid) seed in non-adherent round bottom well plated overnight
- Spheroids were generated and embedded into Matrigel for 16h in precence of CdM
- Images were taken

Tube Formation Assay



Sprouting Spheroid



www.ibidi.com

Angiogenesis models

3. Matrigel plug assay

- Matrigel containing heparin was mixed with DMEM, CdM or exosomes (100 µg/plug)
- C57BL6 mice were anesthetized and then subcutaneously injected with Matrigel along the abdominal midline
- After 2 weeks: animals were sacrificed

Non-contact cell co-culture

- HUVECs were seeded onto the bottom of the plate
- MSCs were seeded and pre-cultured onto the insert (Corning Transwell; membrane cell culture insert)
- Next day:
 - insert was placed into the plate pre-cultured with HUVECs
 - cultured in serum-free DMEM medium for 48h
- Culture medium was cultured and concentrated 100x

Overexpression and knockdown of miR-30b in MSCs and HUVECs

Overexpression:

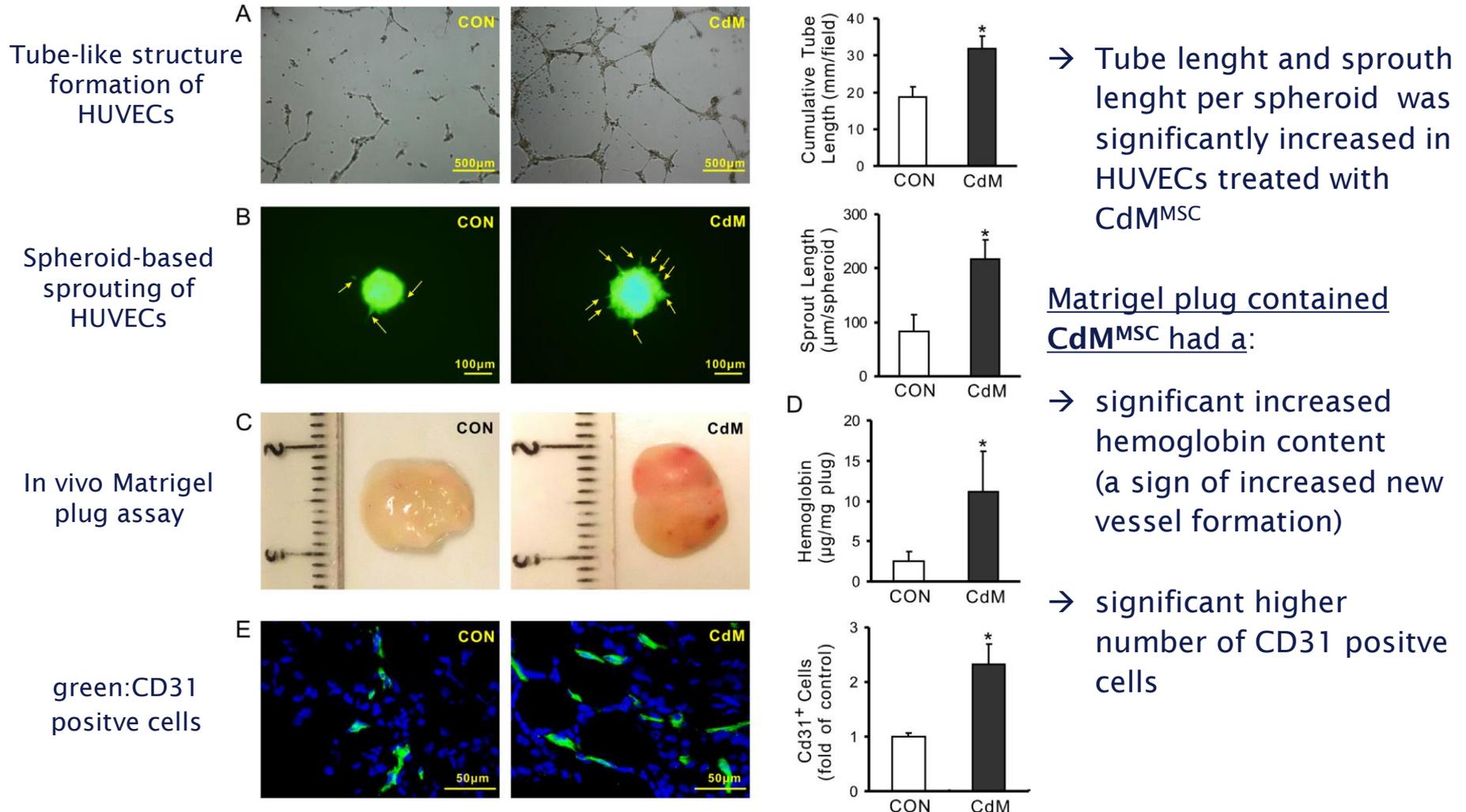
- miR-30b-copGFP expression plasmid and scramble-copGFP control plasmid were co-transfected into 293Ta cells (Lentiviral Packaging Cell Line)
 - for production of high titer lentiviral particles
- Then MSCs and HUVECs were **infected** with high titer lentiviral particles for **24h**

Downregulation

- Synthetic anti-miR-30b was transfected into MSCs using Lipofectamine
- → to downregulate the expression of miR-30b in MSCs

Results

Conditioned medium derived from MSCs promotes angiogenesis



Expression of pro-angiogenic miRNAs in CdM^{MSC} after adding into HUVECs culture for 48hours

miRNA	Downregulated		miRNA	Upregulated	
	CdM ^{MSC} 2 ^(-ΔCt)	CdM ^{MSC} with HUVECs 2 ^(-ΔCt)		CdM ^{MSC} 2 ^(-ΔCt)	CdM ^{MSC} with HUVECs 2 ^(-ΔCt)
miR-424 [#]	44.965 ± 5.542	10.725 ± 1.795*	miR-21	89.021 ± 9.117	187.956 ± 27.620*
miR-30c	6.420 ± 0.623	0.572 ± 0.140*	miR-10a	0.435 ± 0.040	10.160 ± 0.985*
miR-30b	5.877 ± 0.692	0.133 ± 0.012*	miR-126	0.045 ± 0.014	6.988 ± 0.933*
let-7f	4.592 ± 0.245	0.153 ± 0.003*	miR-10b	0.008 ± 0.002	5.869 ± 0.442*
			miR-19a	1.623 ± 0.063	3.380 ± 0.316*
			miR-19b	1.540 ± 0.116	2.950 ± 0.225*

(**P* < 0.05 vs CdM^{MSC}).

[#]The mouse homologue of miR-424 sequence from human is miR-322-5p.

→ Expression of **miR-424**, **miR-30c**, **miR-30b** and **let-7f** in CdM^{MSC} was significantly **reduced** after adding into HUVECs culture

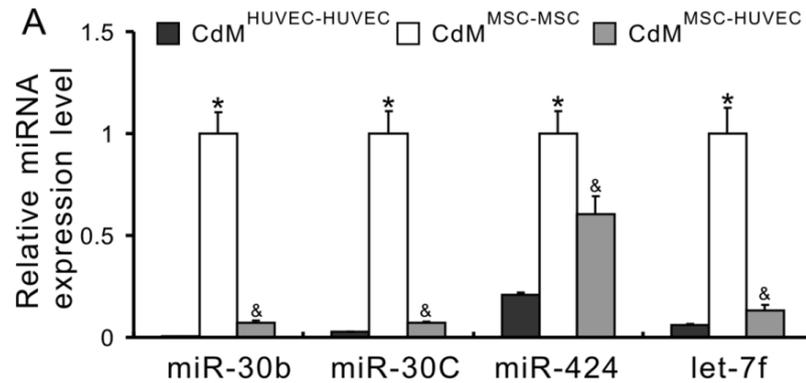
→ indicating that extracellular **miRs transferred into HUVECs**

→ Expression of **miR-21**, **miR-10a**, **miR-126**, **miR-10b**, **miR-19a** and **miR-19b** was significantly **increased** after adding into HUVECs culture

→ Suggesting that **HUVECs might release these miRs**

Transfer of miRNAs between MSCs and HUVECs in a non-contact co-culture system

Supernatant

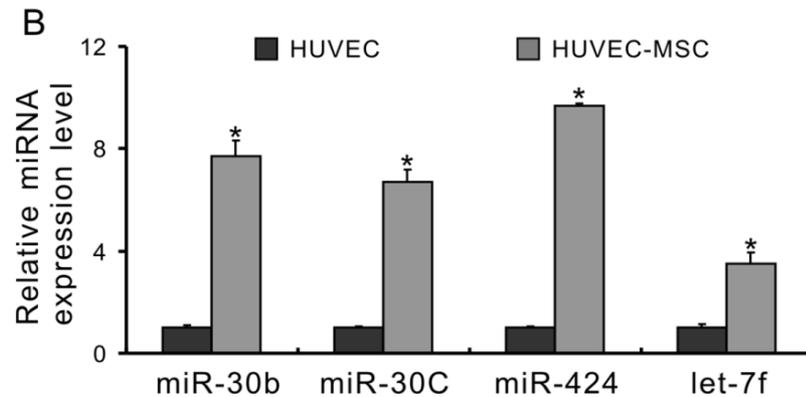


→ The levels of **miR-424, miR-30c, miR-30b** and **let-7f** in

CdM^{HUVEC-HUVEC} was very low (black bars)

CdM^{MSC-MSC} was very high (white bars)

CdM^{MSC-HUVEC} was low (grey bars)



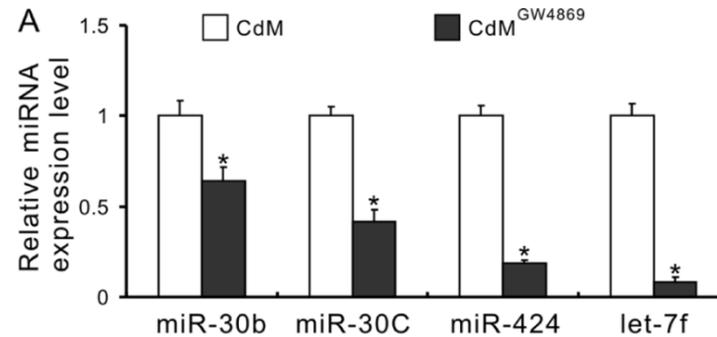
→ The expression of these **miRNAs** in **HUVECs** co-cultured with MSCs was significantly **higher** than in HUVECs without co-cultured with MSC

→ **Demonstrating a transfer of these miRNAs into HUVECs**

Cell lysate

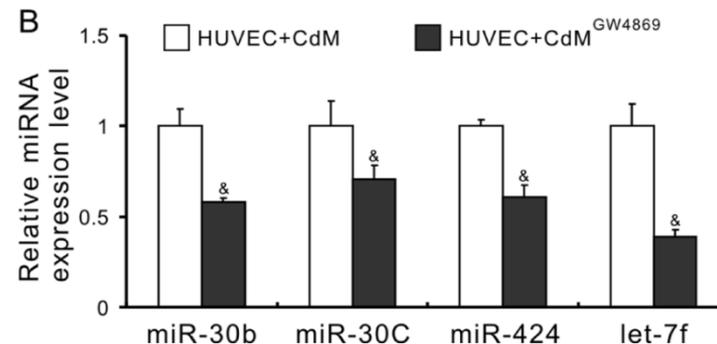
Exosomes derived from MSCs deliver pro-angiogenic miRNAs

Supernatant



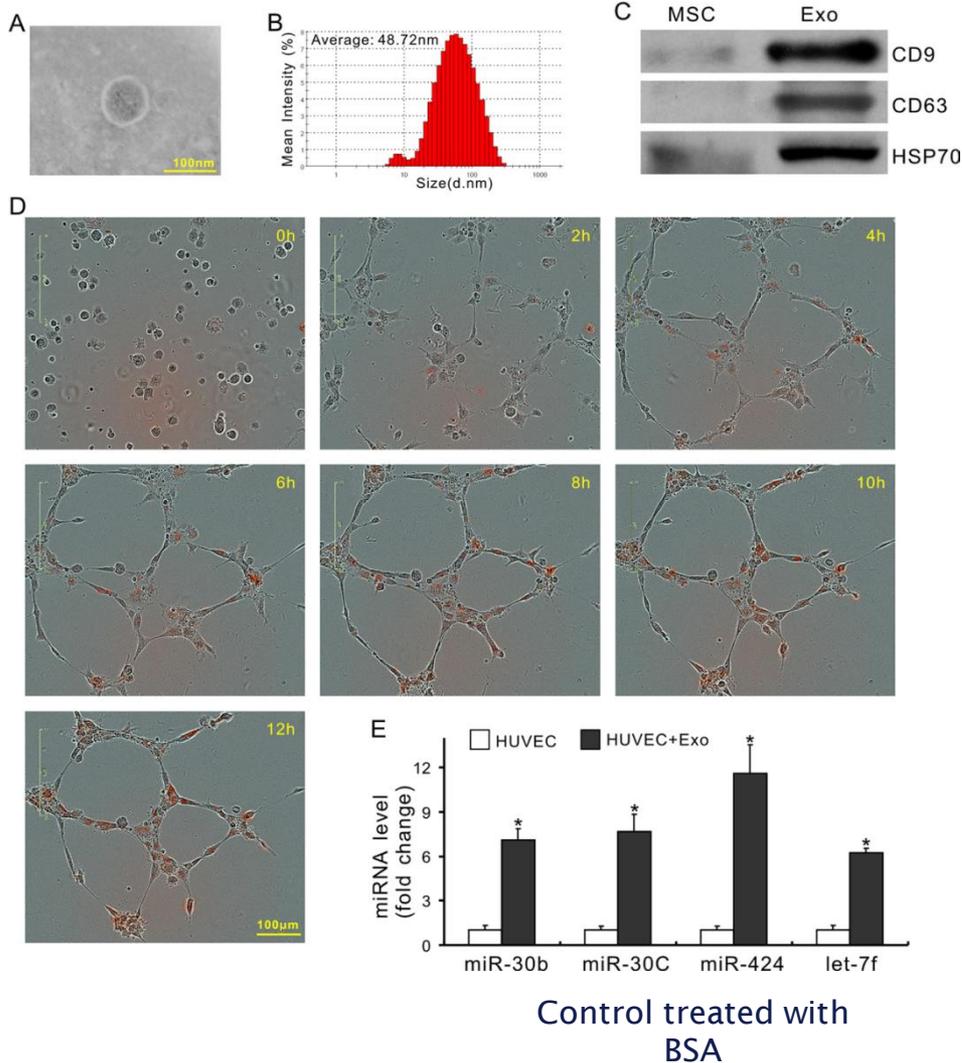
GW4869...an
exosome release
inhibitor

Cell lysate



- The expression of **miR-424**, **miR-30c**, **miR-30b** and **let-7f** in **CdM^{GW4869}** was significantly decreased (A: black bars)
- The levels of these **miRs** in **HUVECs** treated with **CdM^{GW4869}** was significantly reduced (B: black bars)
- **Indicating that exosomes mediated miR transfer between MSC and HUVECs**

Characterization of exosomes derived from MSCs

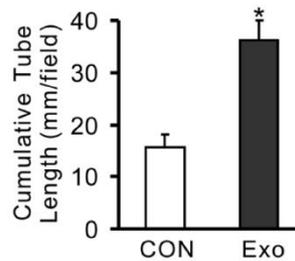
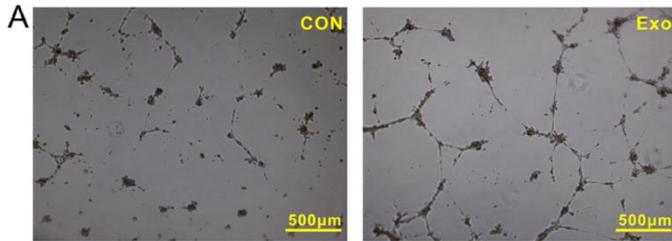


→ Internalization of exosomes pre-labeled with PKH26 (red fluorescence) by HUVECs reached its maximum after 10 h

→ The expression of **miR-424**, **miR-30c**, **miR-30b** and **let-7f** in HUVECs treated with **exosomes** was significantly increased (black bars)

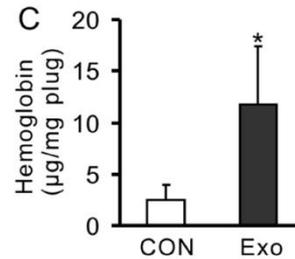
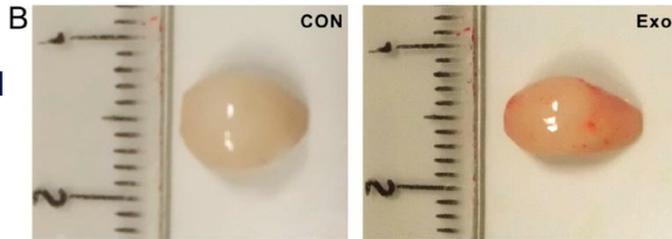
Exosomes derived from MSCs promote angiogenesis

Tube-like structure formation of HUVECs



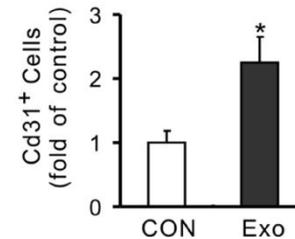
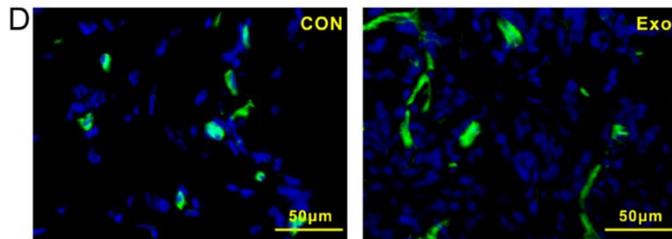
→ Tube length was significantly longer in HUVECs treated with exosomes

In vivo Matrigel plug assay



Matrigel plug contained exosomes had a:

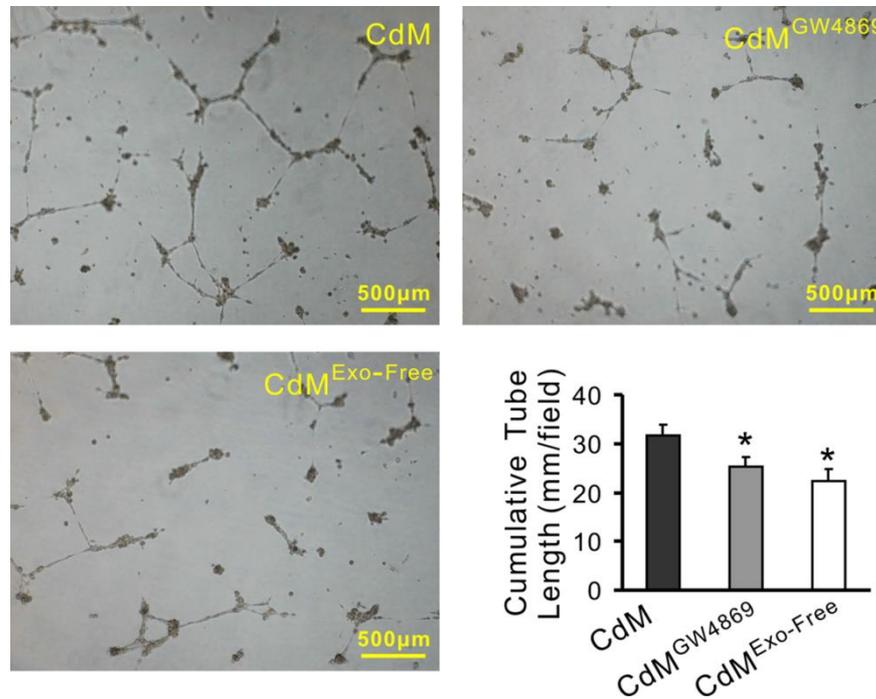
→ significant increased hemoglobin content (a sign of increased new vessel formation)



→ significant higher number of CD31 positive cells

Control treated with BSA (same protein amount)

Exosomes derived from MSCs promote angiogenesis

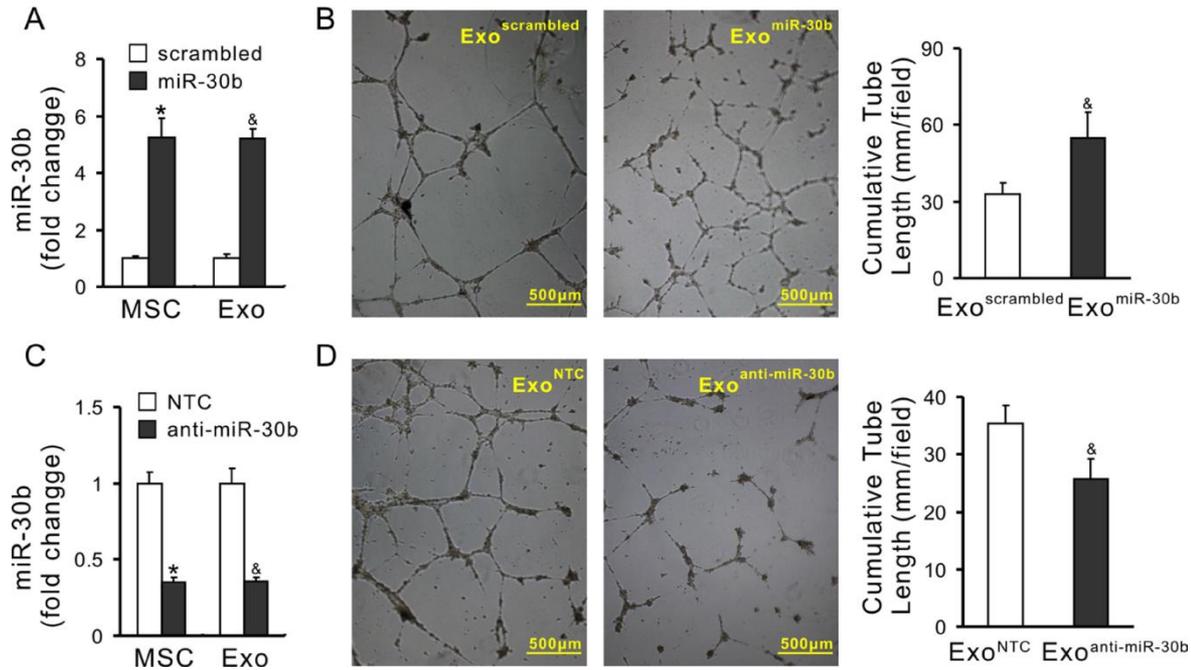


→ Pro-angiogenic capacity of CdM^{MSC} was reduced after inhibiting or depleting exosomes in the CdM

Pro-angiogenic properties of exosomes

Overexpression
of miR-30b in
MSCs using
lentiviral system

Knockdown
of miR-30b
using anti-miR-
30b in MSCs



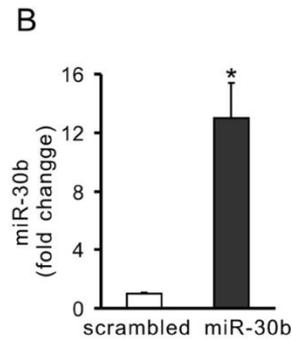
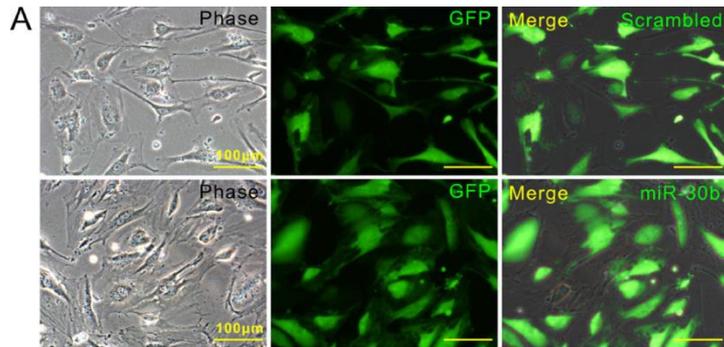
- Expression of miR-30b in **MSC^{miR-30b}** and **Exo^{miR-30b}** was increased
- Tube length was increased in HUVECs treated with Exo^{miR-30b}

- Expression of miR-30b in **MSC^{anti-miR-30b}** and **Exo^{anti-miR-30b}** was reduced
- Tube length was reduced in HUVECs treated with Exo^{anti-miR-30b}

→ Indicating that **overexpression of miR-30b enhanced** and **downregulation of miR-30b reduced** the pro-angiogenic capacity of exosomes

Pro-angiogenic properties of exosomes

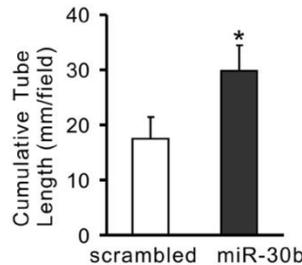
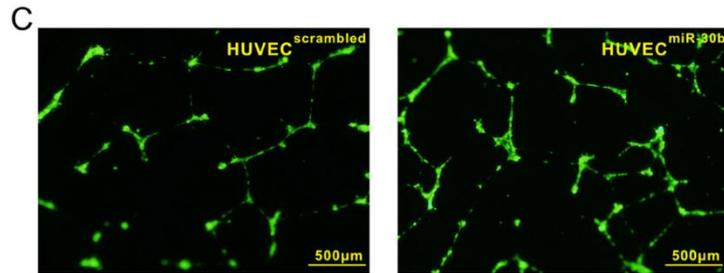
Overexpression of miR-30b in HUVECs using lentiviral system



→ Increased expression of miR-30b and tube length in **HUVECs^{miR-30b}**

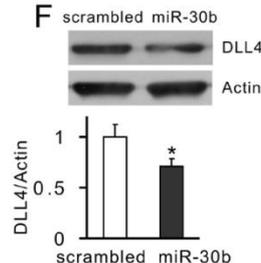
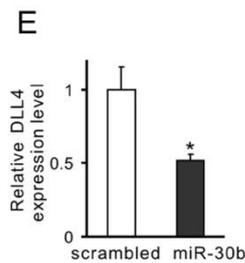
→ TargetScan shows that the 3' UTR of DLL4 contains the conserved miR-30 family binding sites

→ Expression of DLL4 in **HUVECs^{miR-30b}** was significantly reduced



D

	predicted consequential pairing of target region (top) and miRNA (bottom)
Position 59-66 of DLL4 3' UTR	5' ...ACCUUCUUCUGCAUUGUUACA...
hsa-miR-30a	3' GAAGGUCAGCCUCAAUAUGU
Position 59-66 of DLL4 3' UTR	5' ...ACCUUCUUCUGCAUUGUUACA...
hsa-miR-30b	3' UCGACUCACAUCCUACAAUGU
Position 59-66 of DLL4 3' UTR	5' ...ACCUUCUUCUGCAUUGUUACA...
hsa-miR-30c	3' CGACUCUCACAUCCUACAAUGU
Position 59-66 of DLL4 3' UTR	5' ...ACCUUCUUCUGCAUUGUUACA...
hsa-miR-30d	3' GAAGGUCAGCCUCAAUAUGU
Position 59-66 of DLL4 3' UTR	5' ...ACCUUCUUCUGCAUUGUUACA...
hsa-miR-30e	3' GAAGGUCAGUUCUCAAUAUGU



Discussion

- Conditioned medium of MSCs significantly increased tube-like structure formation, spheroid-based sprouting and neo-angiogenesis in Matrigel plug
- Exosomes derived from MSCs:
 - mediated the transfer of miRs from MSCs to HUVECs
 - promoted angiogenesis
- Gain-and-loss function of miRs in exosomes:
 - pro-angiogenic effect is dependent on their pro-angiomiRs cargo

- **MSCs promote angiogenesis through paracrine mechanisms**
- **Angiogenetic effects of MSCs may be related to the secretion of pro-angiomiRs and transfer of these miRs into endothelial cells**
- **Angiogenic effect of CdM was at least partly attributable to exosomes**
- **miR-30b carried by exosomes plays an important role in MSCs mediated angiogenesis**
- **Exosomes contain many growth factors, cytokines and chemokines, which may also participate in angiogenesis**

→ MSC-derived exosomes could be considered for using in therapeutic angiogenesis especially for ischemic diseases

References

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